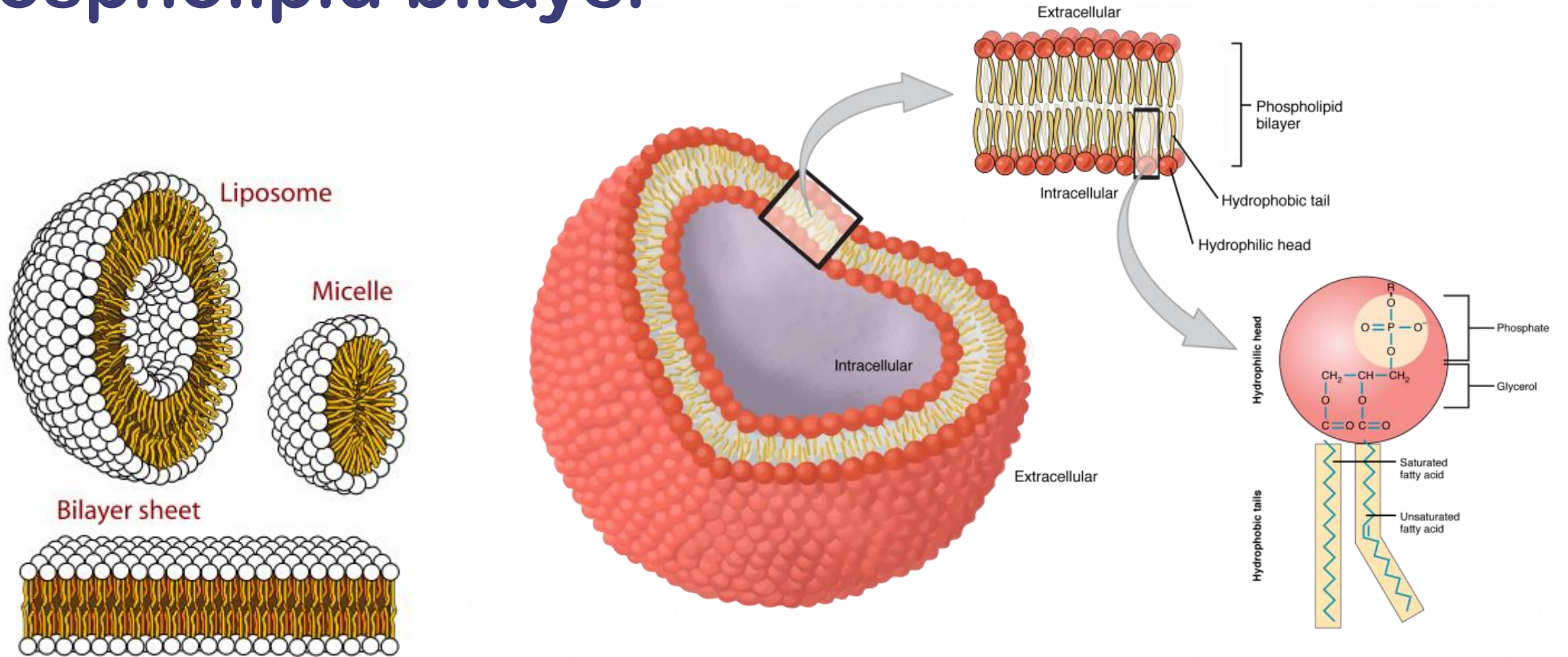


Macromolecules

Transmembrane proteins

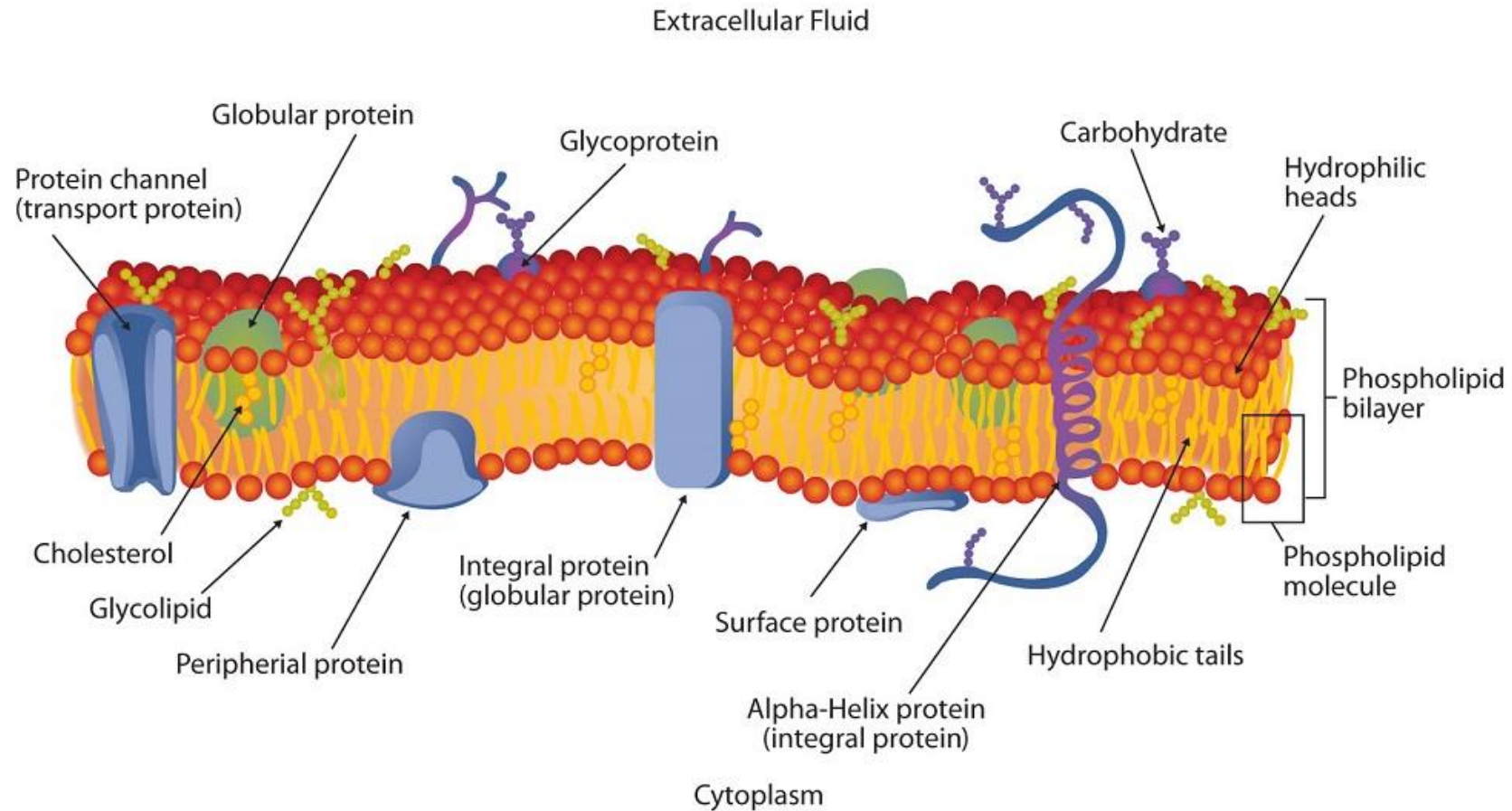
**Csaba Magyar, Institute of Enzymology,
Research Centre for Natural Sciences
2019 December**

Phospholipid bilayer



Separation
Polarity

Function of lipid bilayers



Transmembrane proteins

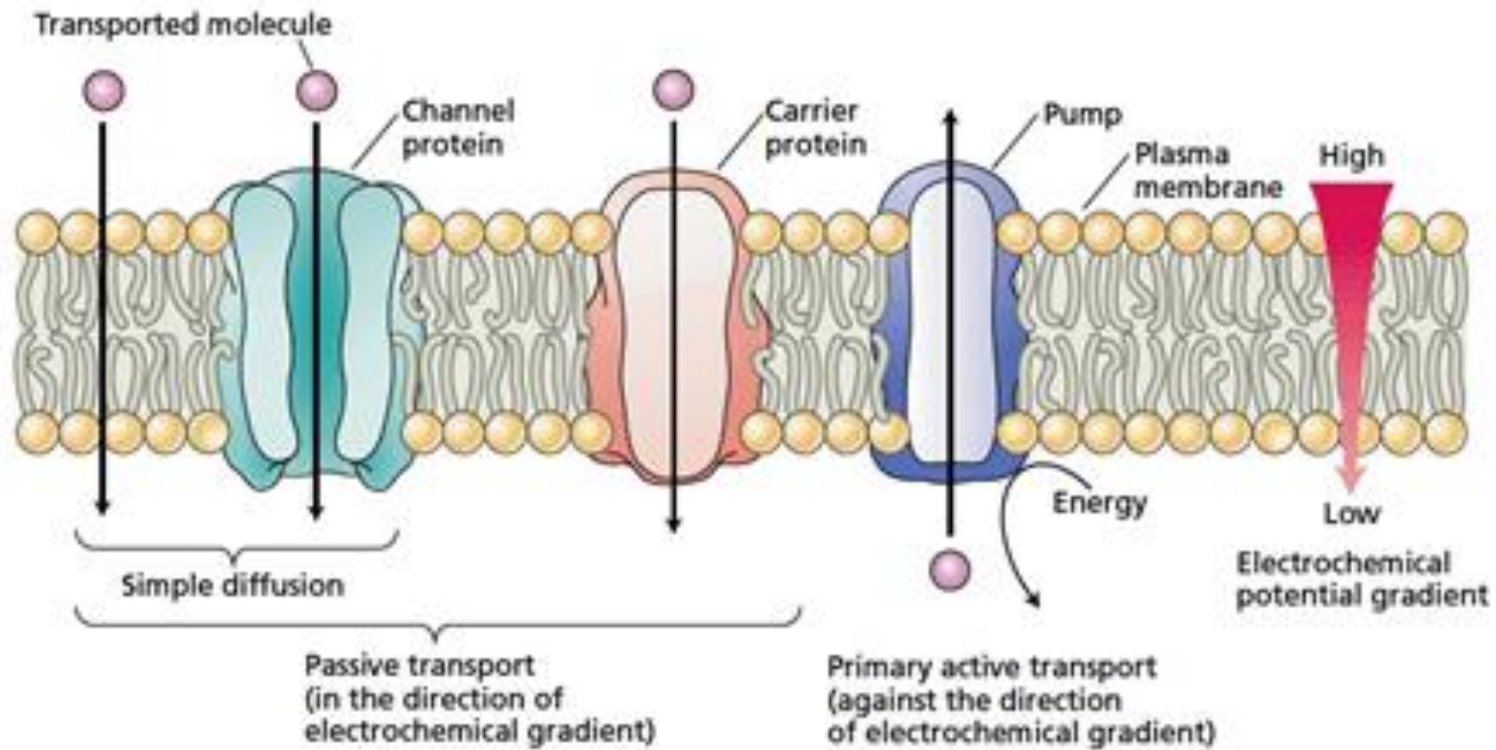
- Lipid bilayer is impermeable to most molecules
- Gates are needed – TM proteins

Transport

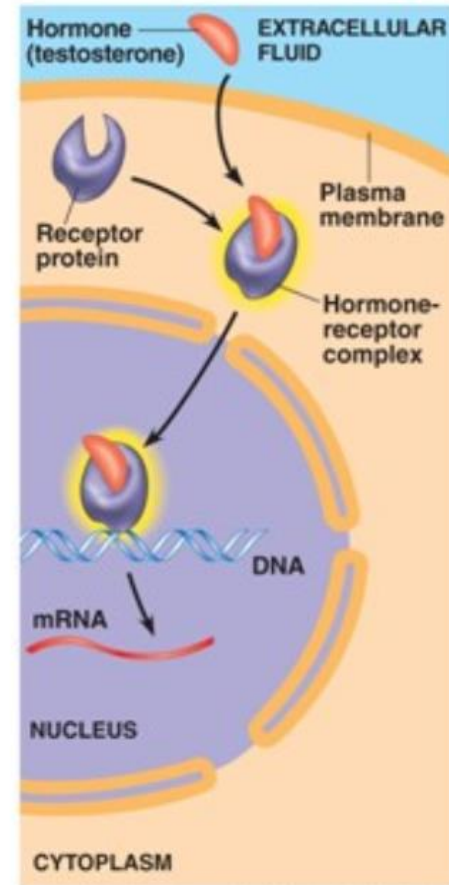
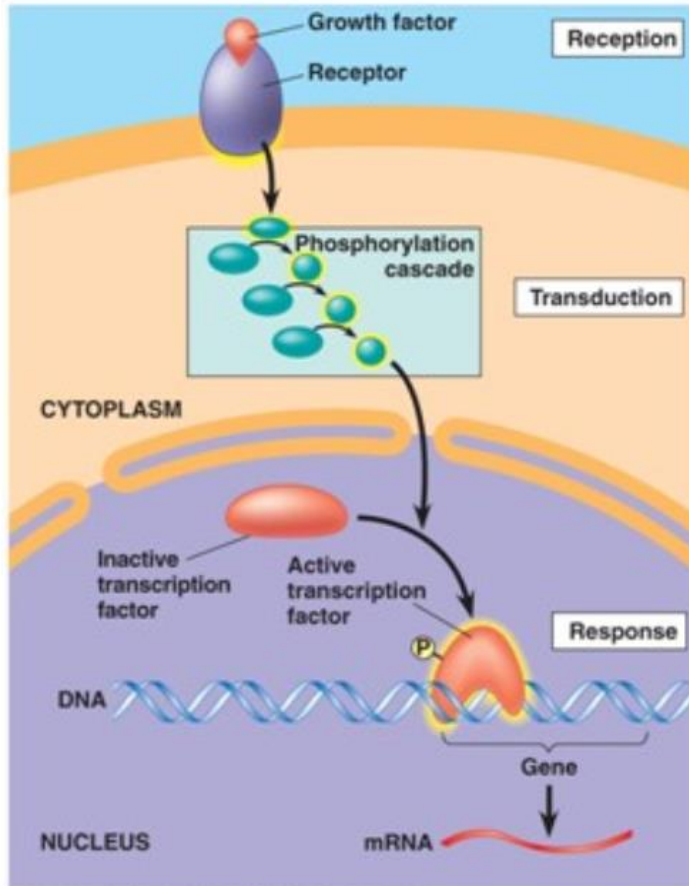
Signal transduction

Energy conversion

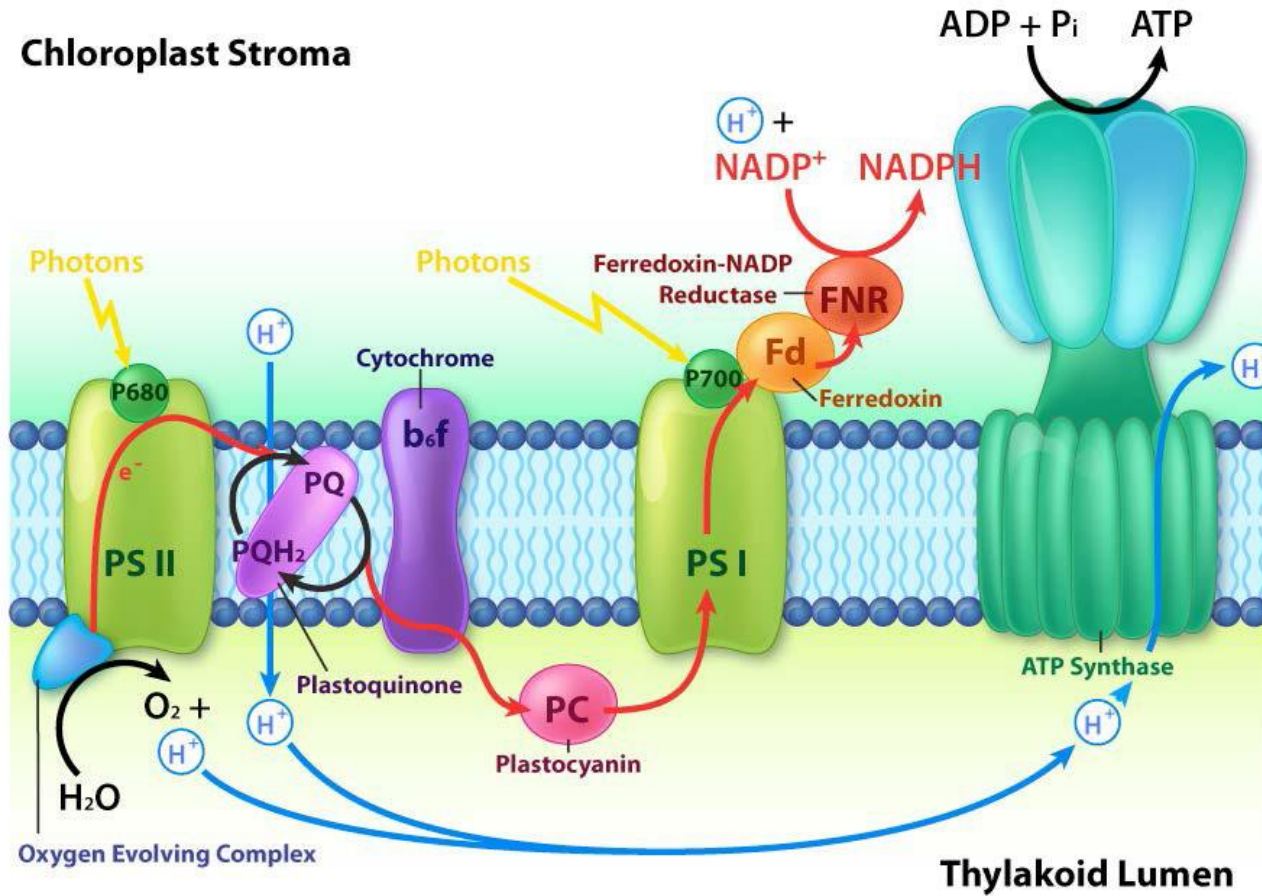
Transport



Signal transduction

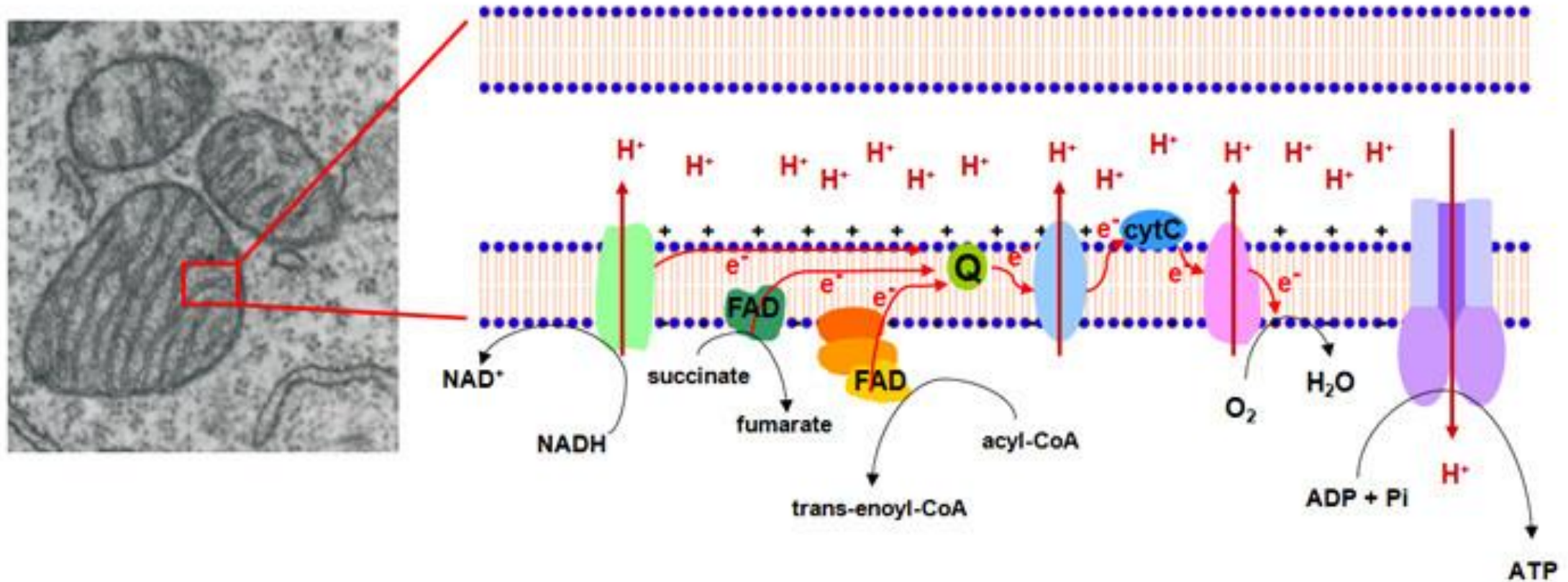


Photosynthesis



Mitochondrion

Krebs cycle, aerobic respiration



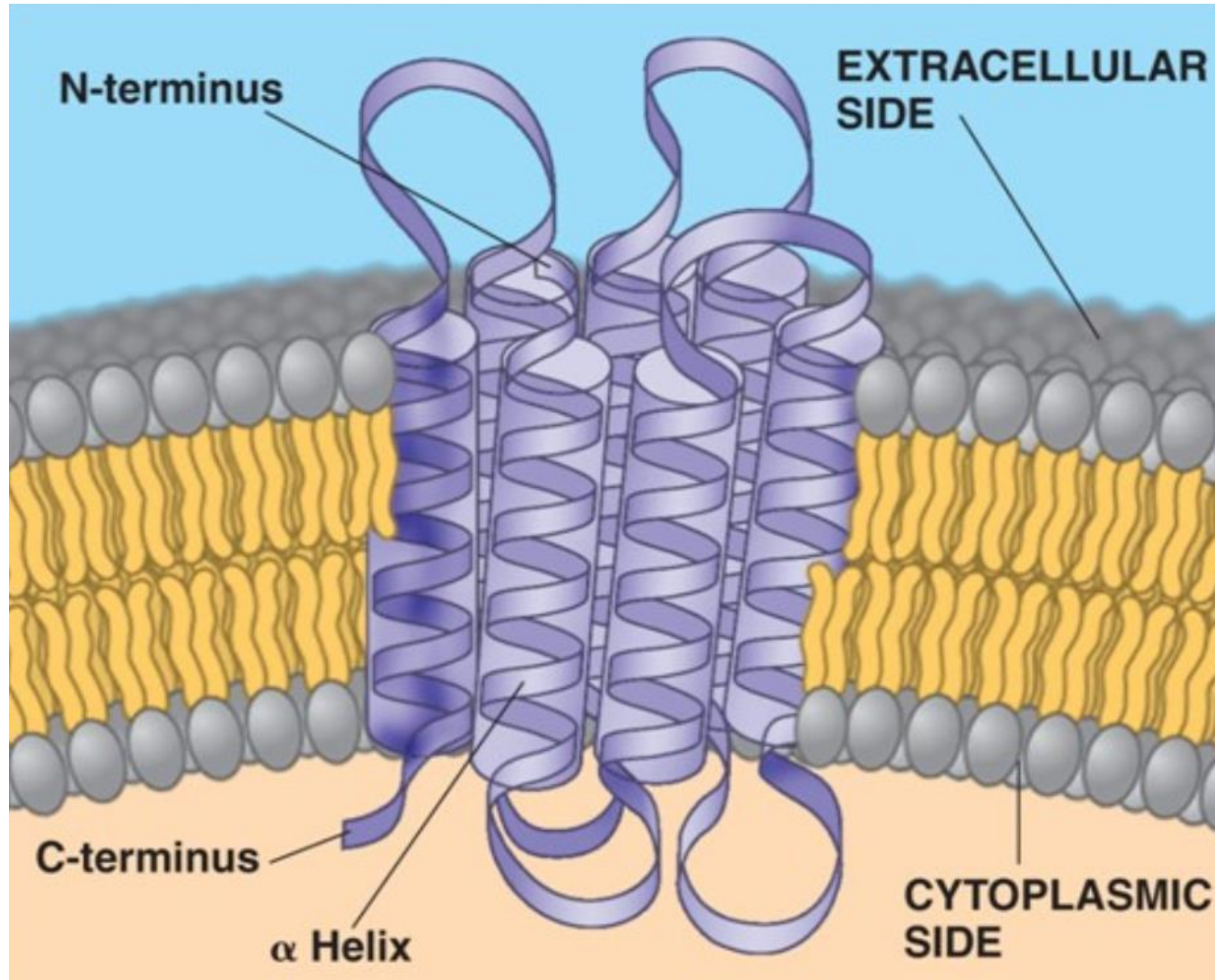
Occurrence

- **Bacteria 1-2% of genome**
- **Eukaryotes ~25%**
 - important drug targets; 134 GPCR targets
 - at least 35% of all drug targets are GPCRs
- **PDB: 2%; underrepresented; hard to crystallize**

Structure

- Alpha
- Beta

Structure is needed
to understand
their function



Structure

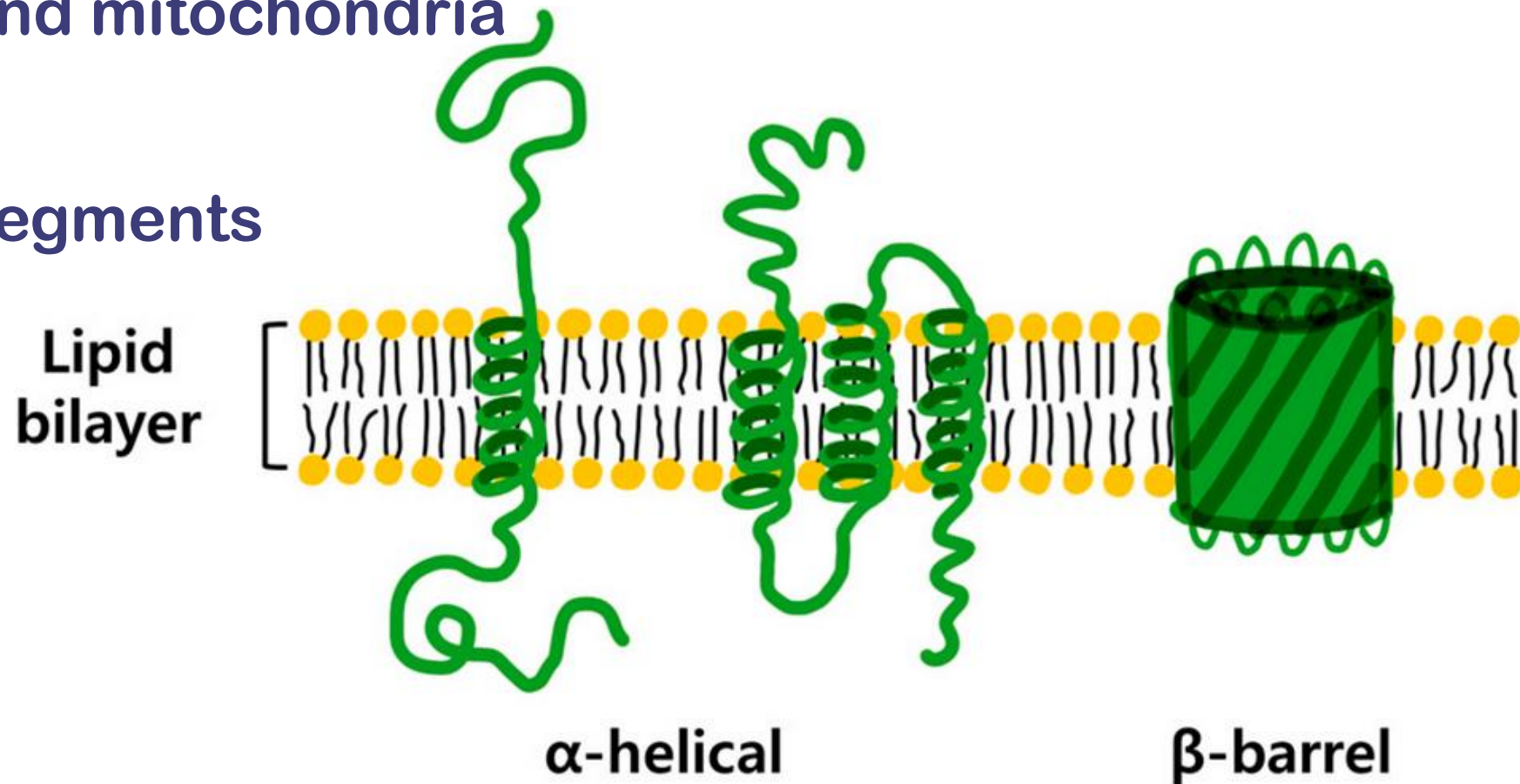
Alpha: from unicellular to mammals

Beta: bacteria and mitochondria

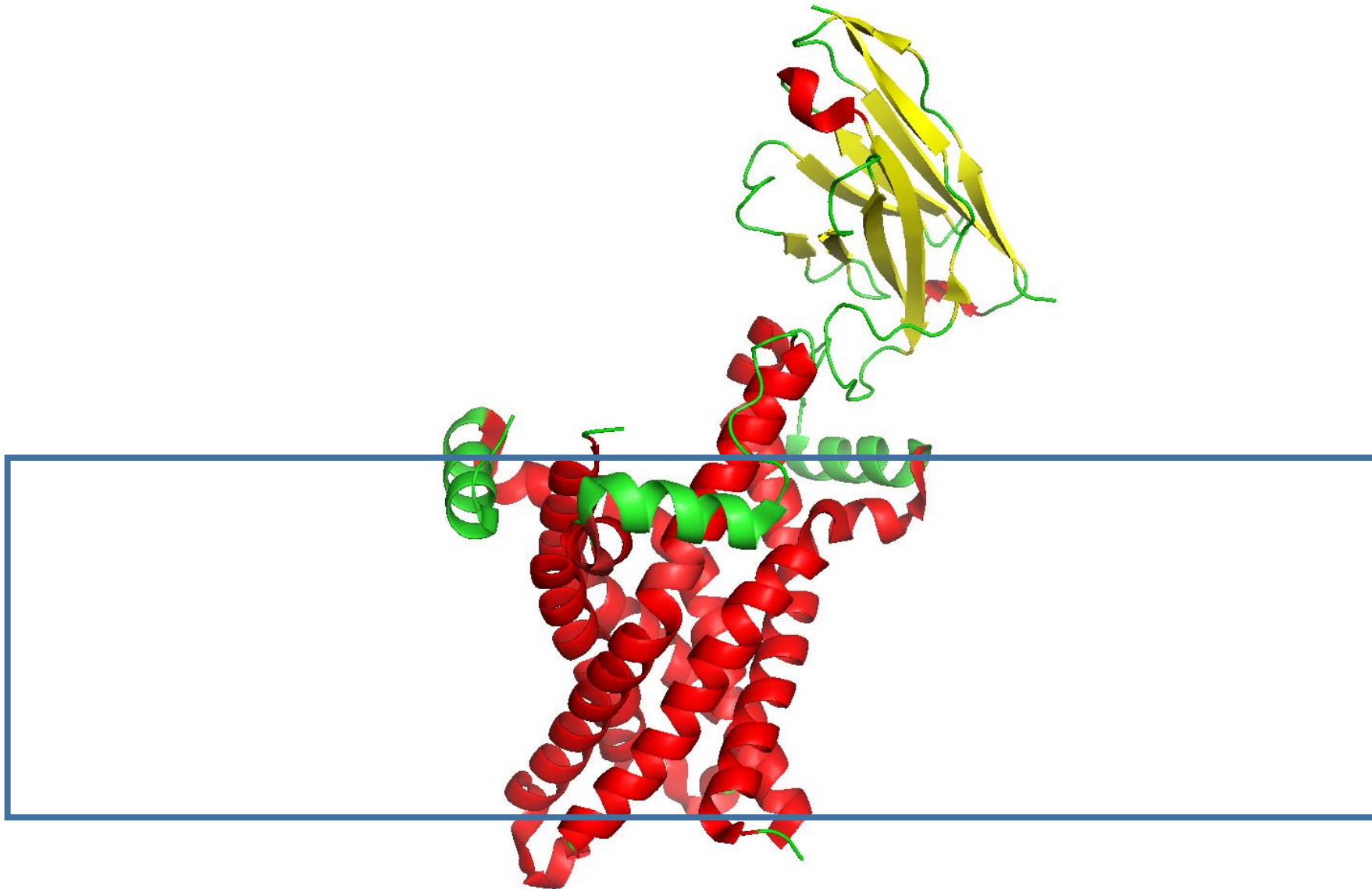
Number of TM segments

1: 40%

6,7: frequent



Orientation of TM helices



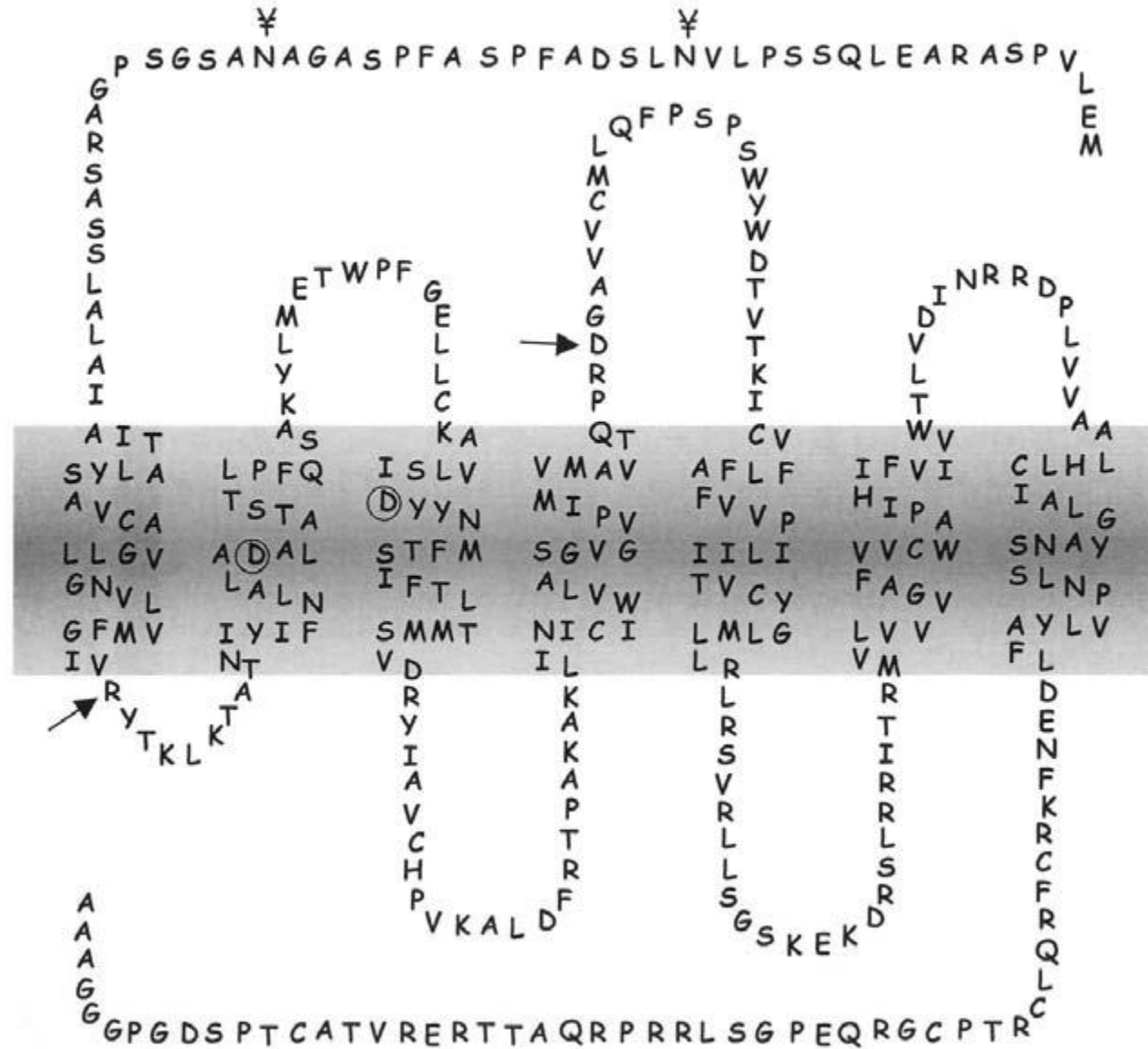
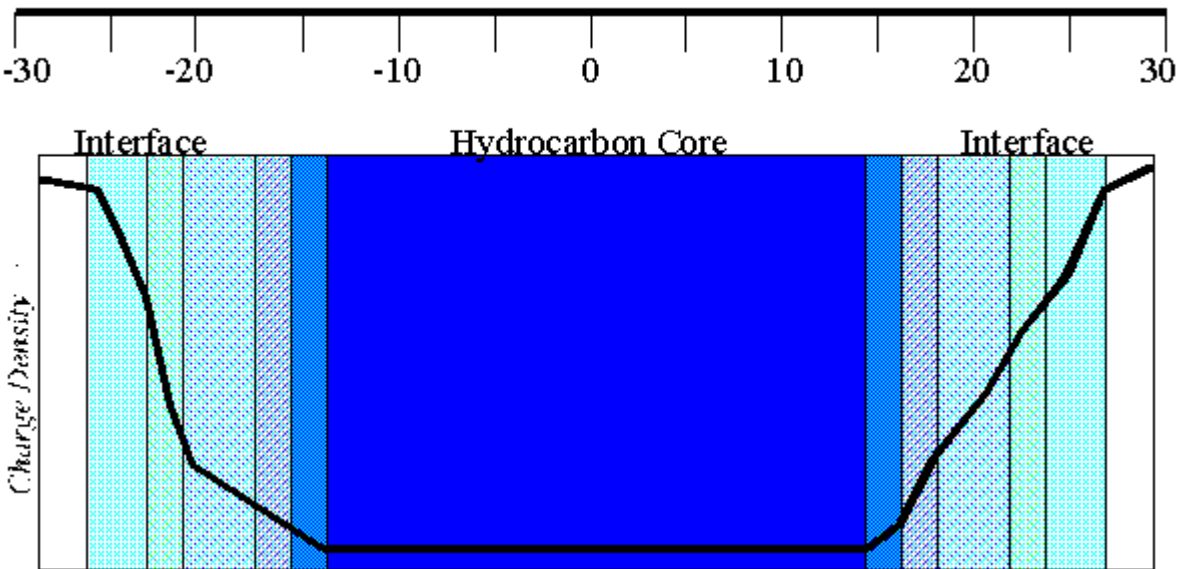
Topography

- Position of TM regions

Size of TM regions

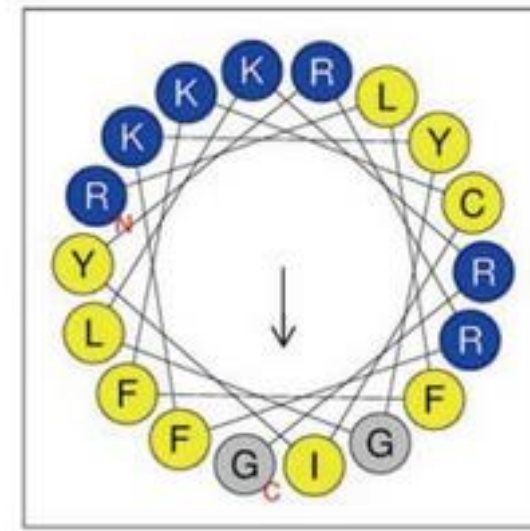
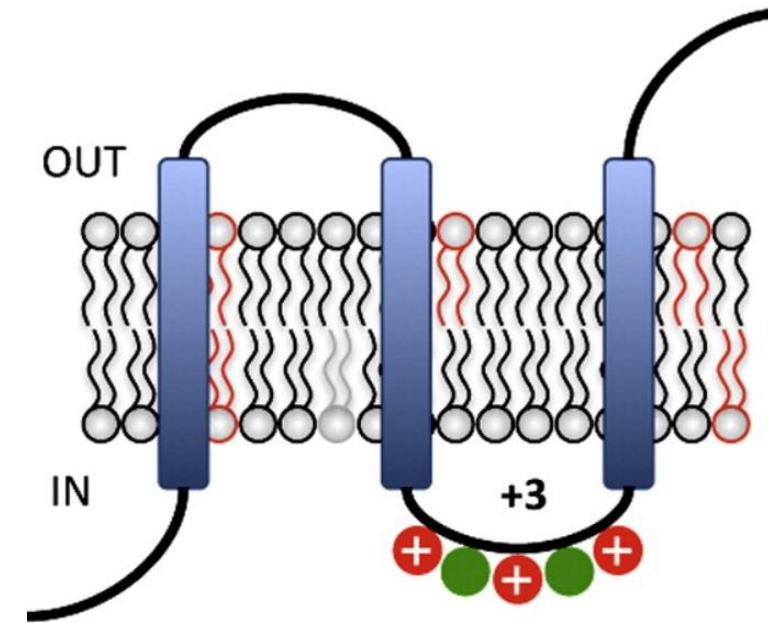
30 Å ~ 20-25 aa

Distance from Hydrocarbon Center (Å)



TM sequences

- Hydrophobic residues, low complexity
- Structure is stabilized with internal H-bond
- Positive inside rule
- Amphipathic helices; anchors
- Proline is a helix breaker, but present in TM helices
missing amide hydrogen - > partial charge
important for function; ion transport



PDBTM database

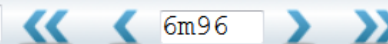


PDBTM: Protein Data Bank of Transmembrane Proteins

PDBTM version: 2019-03-15

Number of transmembrane proteins: 4115 (alpha: 3662 , beta: 429)

all



Home



Search



Download



Statistics



Documents



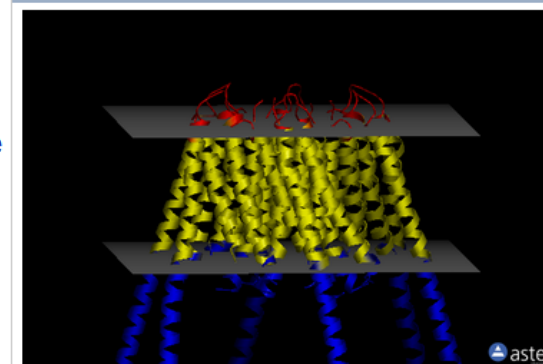
Help



Welcome to the PDBTM home page

PDBTM is the first comprehensive and up-to-date transmembrane protein selection of the Protein Data Bank (PDB). PDBTM database is maintained at the Institute of Enzymology by the Membrane Protein Bioinformatics Research Group. The PDBTM database was created by scanning all PDB entries with the TMDet algorithm. You can get more information about PDBTM in our articles and in the PDBTM manual. If you find PDBTM useful in your research, please cite our articles (Bioinformatics 20, 2964-2972; Nucleic Acids Research 33 Database Issue, D275-D278; Nucleic Acids Research 41 Database Issue, D524-D529).

6aki



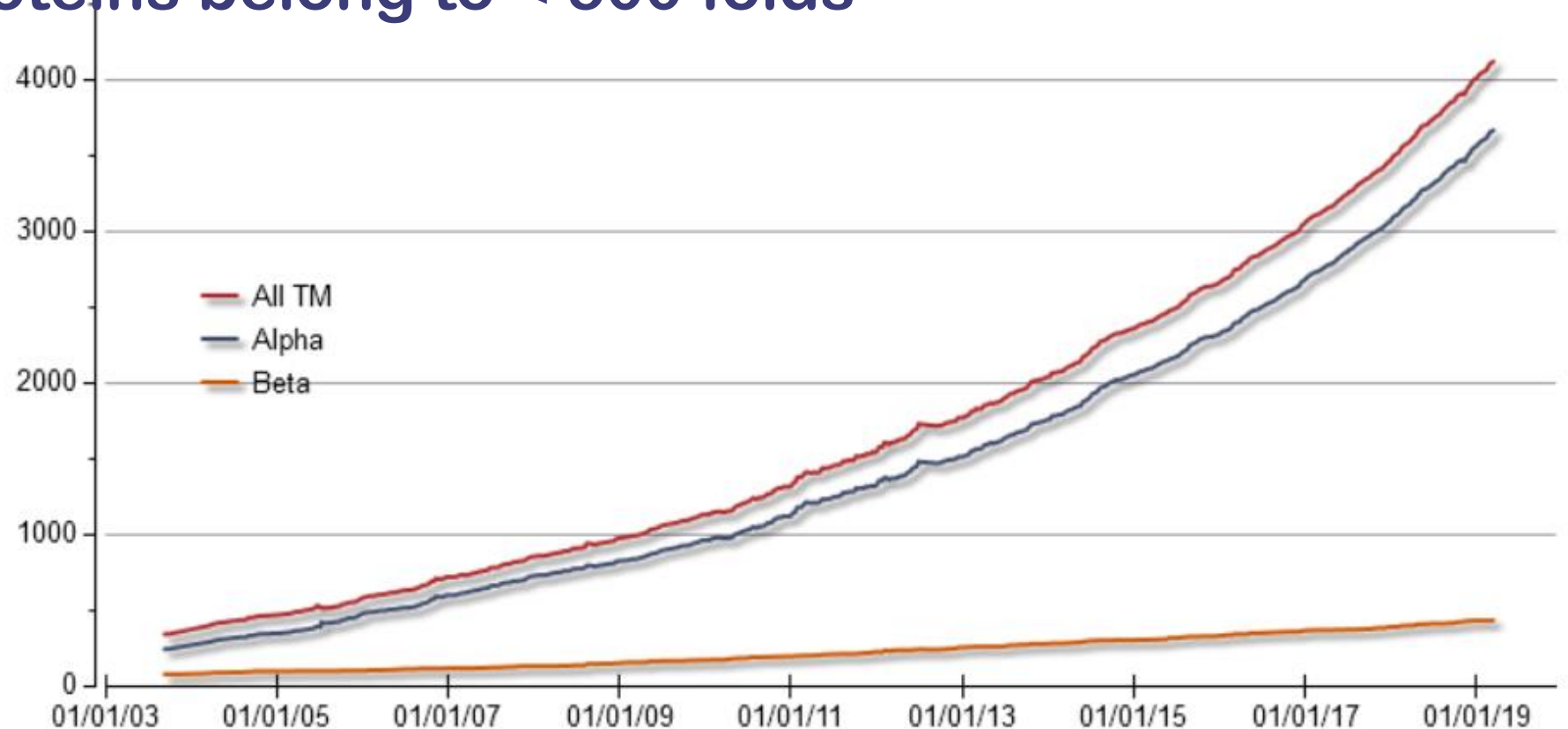
PDBTM type: Tm_Alpha
Chain(s): A[4], B[4], C[4], D[4], E[4], F[4]

TM folds

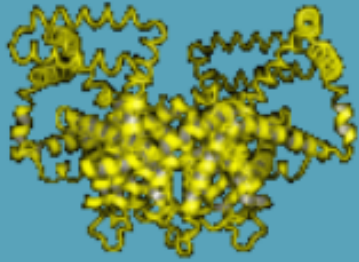
Globular folds < 10k

Large sizes, but small variations

90% of TM proteins belong to < 500 folds



TMDDET



Detection of transmembrane regions by using 3D structure of proteins

Version 2.0

Mon 02 Dec, 2019

General info

TMDDET home page



Form

TMDDET may be used for the detection of the transmembrane regions of membrane proteins using their 3D structure only. You can read more about the algorithm in our [articles](#). You can send your protein structure by using [this form](#).

Manual

Faq

Articles

PDBTM

Comment

If you would like to know the membrane topology of a protein already deposited in the [Protein Data Base](#), please use [PDB_TM](#) database.

OPM

orientations of (OPM) database
proteins in membranes

UNIVERSITY OF MICHIGAN | COLLEGE OF PHARMACY | LOMIZE GROUP

Search proteins by PDB ID or name

[HOME](#) [ABOUT OPM](#) [SEARCH](#) [DOWNLOAD OPM FILES](#) [CONTACT US](#) [PPM SERVER](#) [TMPFOLD SERVER](#)

Orientations of Proteins in Membranes (OPM) database

OPM provides spatial arrangements of membrane proteins with respect to the hydrocarbon core of the lipid bilayer.

OPM includes all unique experimental structures of transmembrane proteins and some peripheral proteins and membrane-active peptides ([Features](#)).

Each protein is positioned in a lipid bilayer of adjustable thickness by minimizing its transfer energy from water to the membrane ([Methods](#)).

OPM provides structural classification and sorting according to different criteria ([Classification](#)).

Our calculations are in agreement with experimental studies of 24 transmembrane and 39 peripheral peptides and proteins.

OPM also provides a few preliminary results of our computational analysis of transmembrane α -helix association in experimental structures of selected polytopic proteins (Assembly pages).

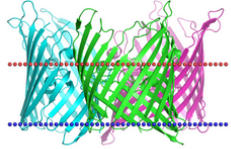
[For more information on single-spanning transmembrane proteins please see our Membranome database](#)

In citing the Orientations of Proteins in Membranes (OPM) database please refer to
Lomize MA, Pogozheva ID, Joo H., Mosberg HI, Lomize AL (2012) OPM database and PPM web server: resources for positioning of proteins in membranes. *Nucleic Acids Res.* **40** (Database issue), D370-D376. [PDF PubMed](#)

For an explanation of our method please refer to
Lomize AL, Pogozheva ID, Lomize MA, Mosberg HI (2006) Positioning of proteins in membranes: A computational approach. *Protein Science.* **15**, 1318-1333. [PDF PubMed](#)

For a new version of our method please refer to
Lomize AL, Pogozheva ID, Mosberg HI (2011) Anisotropic solvent model of the lipid bilayer. 2. Energetics of insertion of small molecules, peptides, and proteins in membranes. *J Chem Inf Model.* **51**, 930-946. [PDF PDF \(supplementary\) PubMed](#)

For more information on peripheral proteins please refer to
Lomize AL, Pogozheva ID, Lomize MA, Mosberg HI (2007) The role of hydrophobic interactions in positioning of peripheral proteins in membranes. *BMC Struct Biol.* **7**, 44. [PDF PubMed](#)



Porin B (PorB), different strain
pdb-3wi4

[Download File: 3wi4.pdb](#)

Structure Statistics (distinct protein structures/PDB entries)

Type: Transmembrane - (2579/5883)
Class: Alpha-helical polytopic - (1850/4069)
Class: Bitopic proteins - (442/1308)
Class: Beta-barrel transmembrane - (287/506)

Type: Monotopic/peripheral - (1513/4986)
Class: All alpha monotopic/peripheral - (320/1207)
Class: All beta monotopic/peripheral - (572/1846)
Class: Alpha/Beta monotopic/peripheral - (338/1304)
Class: Alpha + Beta monotopic/peripheral - (283/629)

Type: Peptides - (647/1159)
Class: Alpha-helical peptides - (448/787)
Class: Beta-hairpin peptides - (125/229)
Class: Beta-helical peptides - (17/36)
Class: Peptides of nonregular structure - (57/107)

Protein Classification

Types (3)
Classes (11)
Superfamilies (504)
Families (976)
Species (854)
Localizations (24)
Proteins (4739)

Assembly

Superfamilies (9)
Families (19)
Localizations (8)
Assemblies (207)

Protein Links

[PDB Sum](#), [PDB](#), [MPKS](#), [MPDB](#)

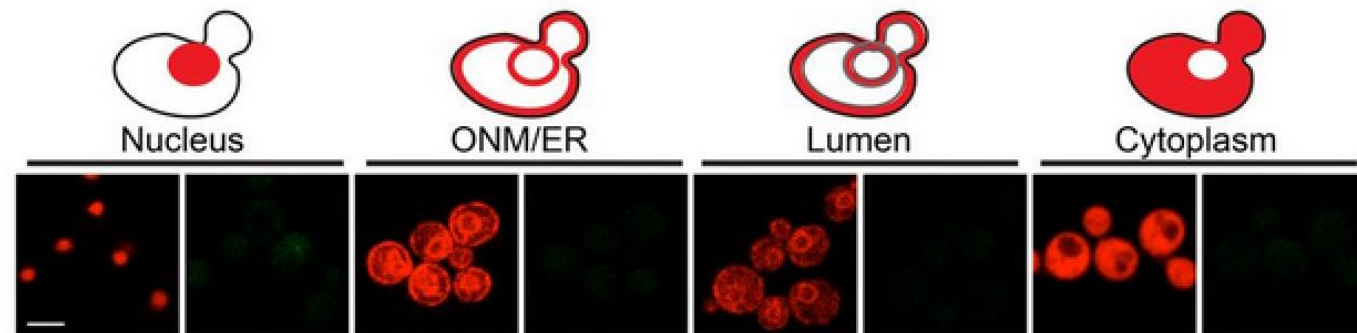
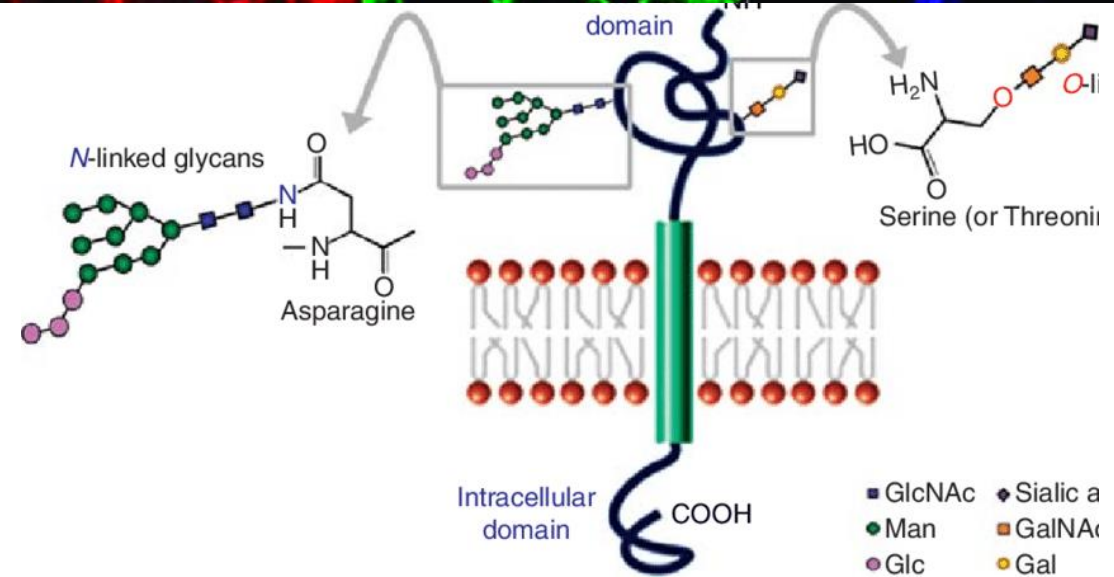
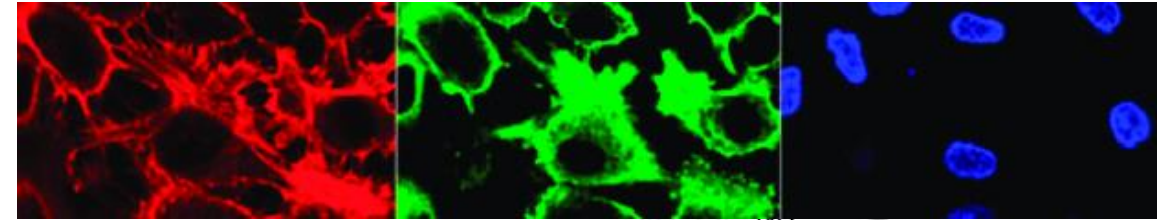
PPM Server

TMPfold Server

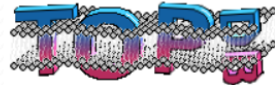
Experimental methods

- Immuno localization; antibodies
- Glycosylation sites
- Reporter enzyme fusion; GFP

Data collected in TOPDB



TOPDB



Topology Data Bank of Transmembrane Proteins *Topology, Structure and Prediction.*

Database revision: (4190 entries, 75211 topology data)

Quick search:

Menu

- ▶ [Home](#)
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- ▶ [Statistics](#)
- ▶ [Contact](#)
- ▶ [Related servers](#)

Database status

Database revision:	
Revision date:	01/01/70
Entries:	4190
Topology data:	75211
Alpha helical proteins:	4067
Beta barrel proteins:	123
PubMed links:	4270
PDB links:	12816
UniProt links:	4190
Visitors:	505773

TOPDB: Topology Database of Transmembrane Proteins

About TOPDB

The Topology Data Bank of Transmembrane Proteins (TOPDB) is currently the most complete and comprehensive collection of transmembrane protein datasets containing experimentally derived topology information. It contains records for 4190 transmembrane proteins with information gathered from the literature and from public databases available on the World Wide Web.

The database collects the details of various experiments carried out to learn about the topology of particular transmembrane proteins. The experimental techniques include fusion with reporter enzymes, glycosylation studies, protease accessibility, immunolocalisation, etc. In addition to literature-derived data, an extensive collection of structural data was also compiled from [Protein Data Bank \(PDB\)](#) and from [Protein Data Bank of Transmembrane Proteins \(PDBTM\)](#) by utilising the [TMDET](#) algorithm.

While literature-derived data can not be collected automatically, data based on 3D structures provides semi-automatic and continuously updated information for the database. Structural data is the most reliable information about transmembrane topologies, but the topology information is often incomplete. Therefore, for each protein in the database the most probable topology consistent with the collected experimental constraints was also calculated using [CCTOP](#) and [HMMTOP](#) transmembrane topology prediction algorithms for α -helical and β -barrel transmembrane proteins, respectively.

Each record in TOPDB also contains the indispensable information about the given protein such as its sequence, name, organism and cross references to various databases ([PDB](#), [PDBTM](#), [UniProt](#) and literature references from [PubMed](#)).

This web interface of TOPDB includes tools for extensive [searching](#), [relational querying](#) and [data browsing](#) as well as [visualisation tools](#) for topology data.

The TOPDB is designed to address the broad gap between the large number of transmembrane proteins in sequence databases and the publicly available topology information of experimentally or computationally studied transmembrane proteins.

Methods use for TM proteins

- Signal peptide identification
- Differentiate TM and globular sequences (DAS-TMfilter)
- Topology prediction (HMMTOP)
- Fold recognition (TOPDOM)
- Homology modelling
- Membrane positioning (TMDET, OPM)

TOPDOM



TOPDOM: Conservatively Located Domains and Motifs in Proteins

[Home](#) [Documents](#) [Download](#) [Search](#) [Related sites](#)

TOPDOM is

a collection of domains and sequence motifs located conservatively in one side of membranes either in transmembrane or globular proteins. The database was created by predicting the transmembrane status and topology of all protein sequence in **UniProt** database by the **CCTOP** algorithm and scanning by specific domain or motif detecting algorithms. The identified domain or motif was added to the database if it was uniformly annotated in the same side of the membrane of the various proteins in **UniProt** database. The sequences in the **UniProt** database were scanned by using hmmpfam algorithm with **Pfam** and **Smart** domain databases, prosite scan with **Prosite** motif database and fingerprints algorithm with **Prints** database. For further details refer to our **manuscripts**.

TOPDOM can be

downloaded either the whole database or various subset of the database in XML format. The database can be searched by **keywords** or **identifiers**. Moreover, users can **scan** their protein sequences against the TOPDOM domains and motifs.

Constrained prediction

can be performed after sequence **scanning** by the **CCTOP** transmembrane topology prediction algorithm using the search results as constrains.

Statistics

Visitors	444186
Entries	6232
Pfam	3930
Prints	589
Prosite	1375
Smart	338
UniProt	195371
Inside	4440
Outside	1792
Transmem	1331
Search by sq	1109268
Search by kw	5679
Search by id	12422

Topology prediction

- Structure determination is difficult
- Prediction methods are needed

Machine learning

Neural networks

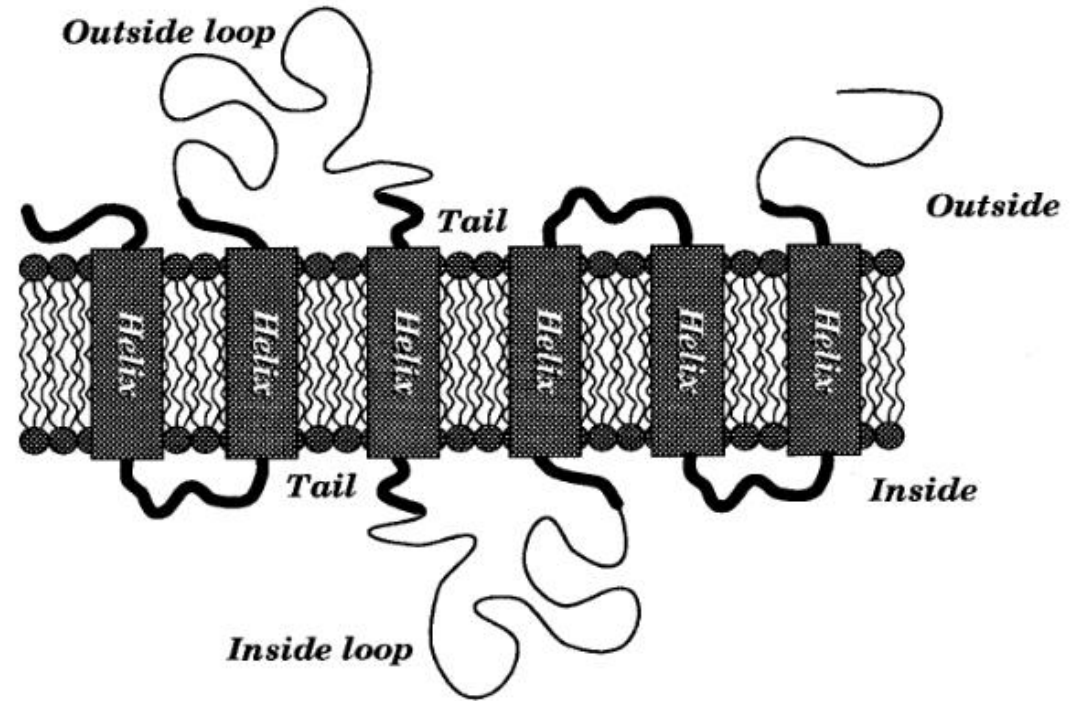
Hidden Markov model

Support Vector Machine

HMMTOP

Hypothesis: the topology is determined by the difference in the amino acid distributions.

o->h->i->h->o hidden Markov model



Amino acid seq: MGDVCDTEFGILVA...SVALRPRKHGRWIV...FWVDNGTEQ...PEHMTKLHMM...

State seq: oooooooooohhhhh...hhhhiiiiiiihhh...hhhooooOO...OOooooohhh...



The most likely topology is the one that maximizes this difference.

Incorporation of experimental data

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Copyright © G. E. Tusnady, 2001

HMMTOP

*Prediction of transmembrane helices
and topology of proteins*

Version 2.0

by G. E. Tusnady



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Copyright © G. E. Tusnady, 2001

Your sequence(s):

Sequence Format: ([help](#)) Unformatted ▾

Sequence type: ([help](#)) Single Sequence(s) ▾

Prediction type: ([help](#)) Reliable ▾

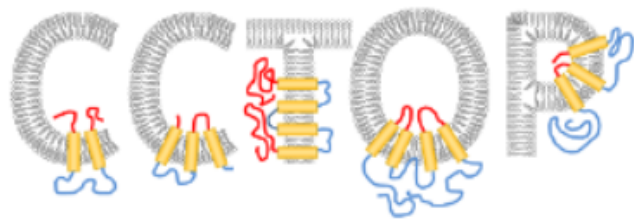
Localization of sequence part(s) ([help](#))

Output format: ([help](#)) HTML ▾

Results in one line: ([help](#))

Submit Clear

CCTOP



Constrained Consensus TOPology prediction server



Documents



Predictions

Visitors:	1206895
Prediction count:	67769
Server version:	s.1.00
Last changed:	13.07.2017

Welcome to the Constrained Consensus TOPology prediction server.

Given the amino acid sequence of a putative α -helical transmembrane protein, CCTOP predicts its topology i.e. localization of membrane spanning regions and orientation of segments between them. The prediction is a consensus of 10 different methods enhanced with available structural and experimental information of any homologous proteins in the [TOPDB](#) database. CCTOP was tested on a benchmark set containing 170 proteins with known structure and achieved the highest accuracy among state-of-art and consensus methods.

You can find more information how CCTOP works in the [description](#) or in the [article](#). A short description about the server can be found in the [manual](#).

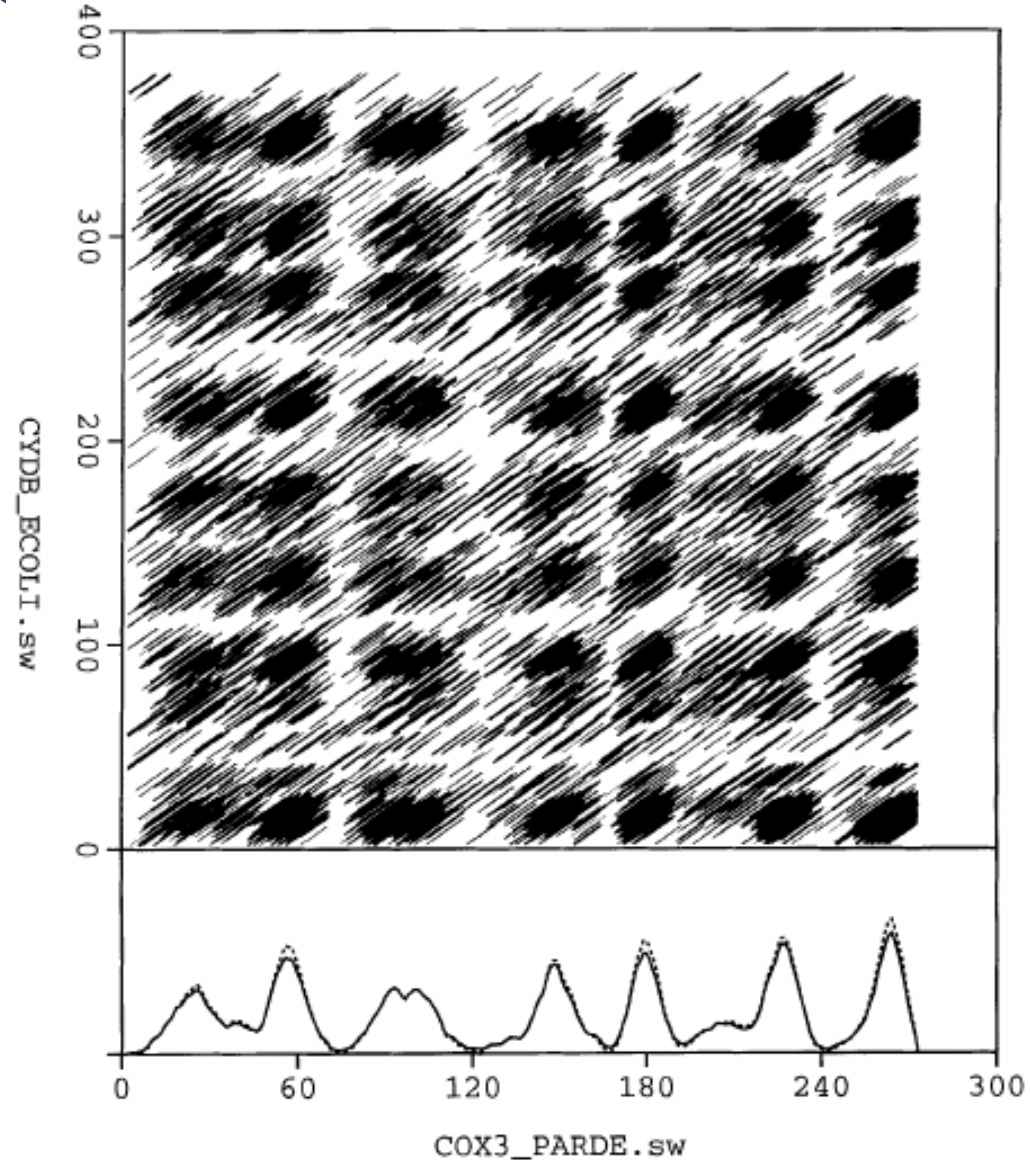
You can submit protein sequences [here](#).

Other methods

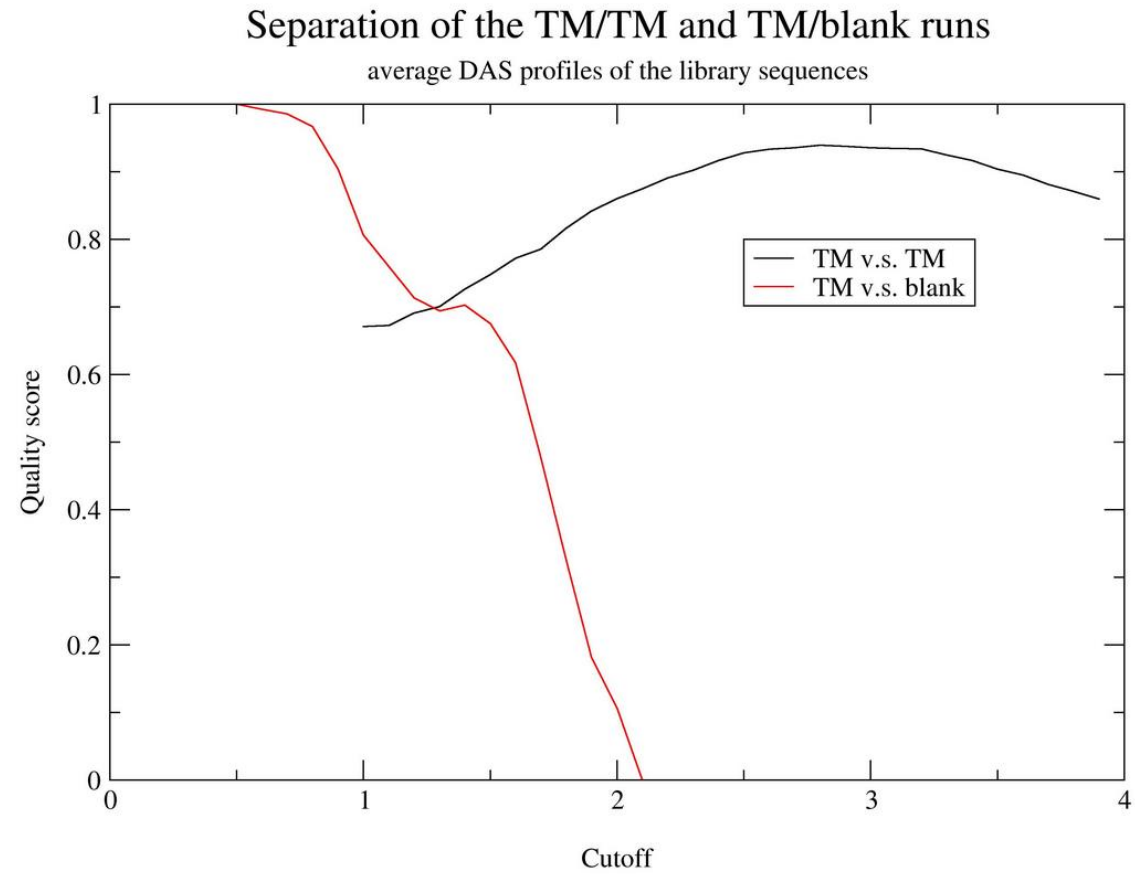
- **DAS** - Prediction of transmembrane regions in prokaryotes using the Dense Alignment Surface method (Stockholm University)
- **MemBrain** - Transmembrane protein structure prediction (HongBin Lab)
- **Memsat** - Membrane Helix Prediction (University College London)
- **Octopus** - Prediction of membrane protein topology and signal peptides (Stockholm University)
- **Philius** - Transmembrane Topology Prediction Server (Yeast Resource Center)
- **Phobius** - A combined transmembrane topology and signal peptide predictor (University of Copenhagen)
- **PredictProtein** - Prediction of transmembrane helix location and topology (Columbia University)
- **Pro and Prodiv** - Alpha-helical transmembrane protein topology prediction methods utilizing hidden Markov models and evolutionary information (Stockholm University)
- **SignalP** - Predicts the presence and location of signal peptide cleavage sites (Technical University of Denmark)
- **Scampi** - Prediction of membrane protein topology from first principles (Stockholm University)
- **SOSUI** - Prediction of transmembrane regions (Nagoya University, Japan)
- **TMHMM** - Prediction of transmembrane helices in proteins (CBS; Denmark)
- **TMpred** - Prediction of transmembrane regions and protein orientation (EMBnet-CH)
- **TopPred** - Topology prediction of membrane proteins (France)

Dense Alignment Surface method (DAS)

Hydrophobic
TM helices
are similar to
each other

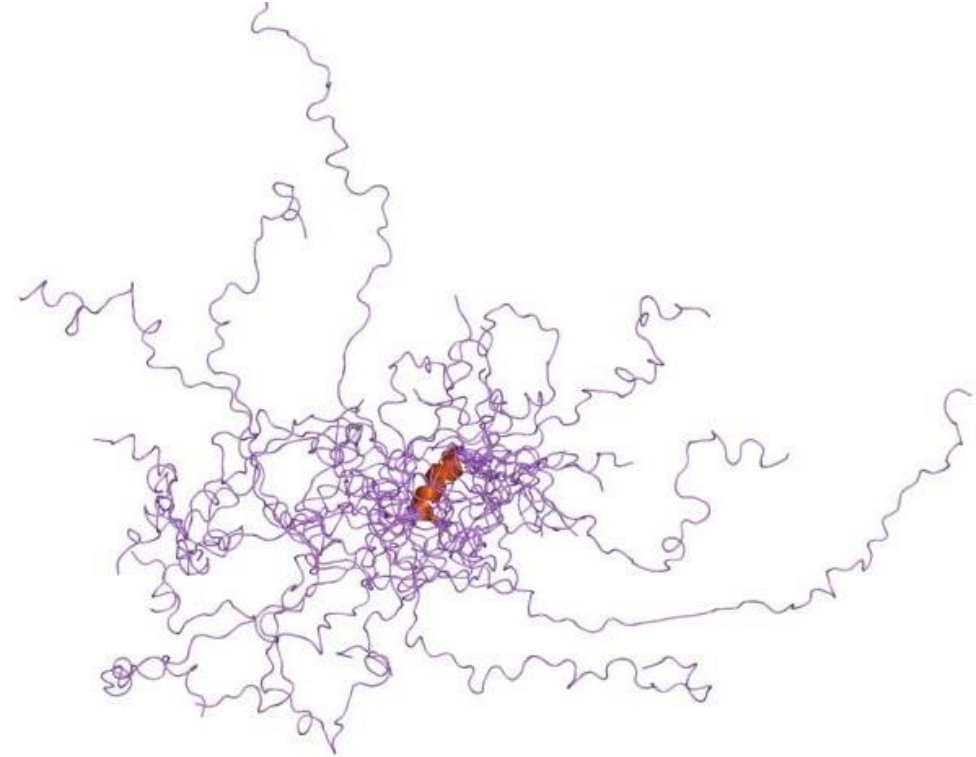


DAS-TMfilter



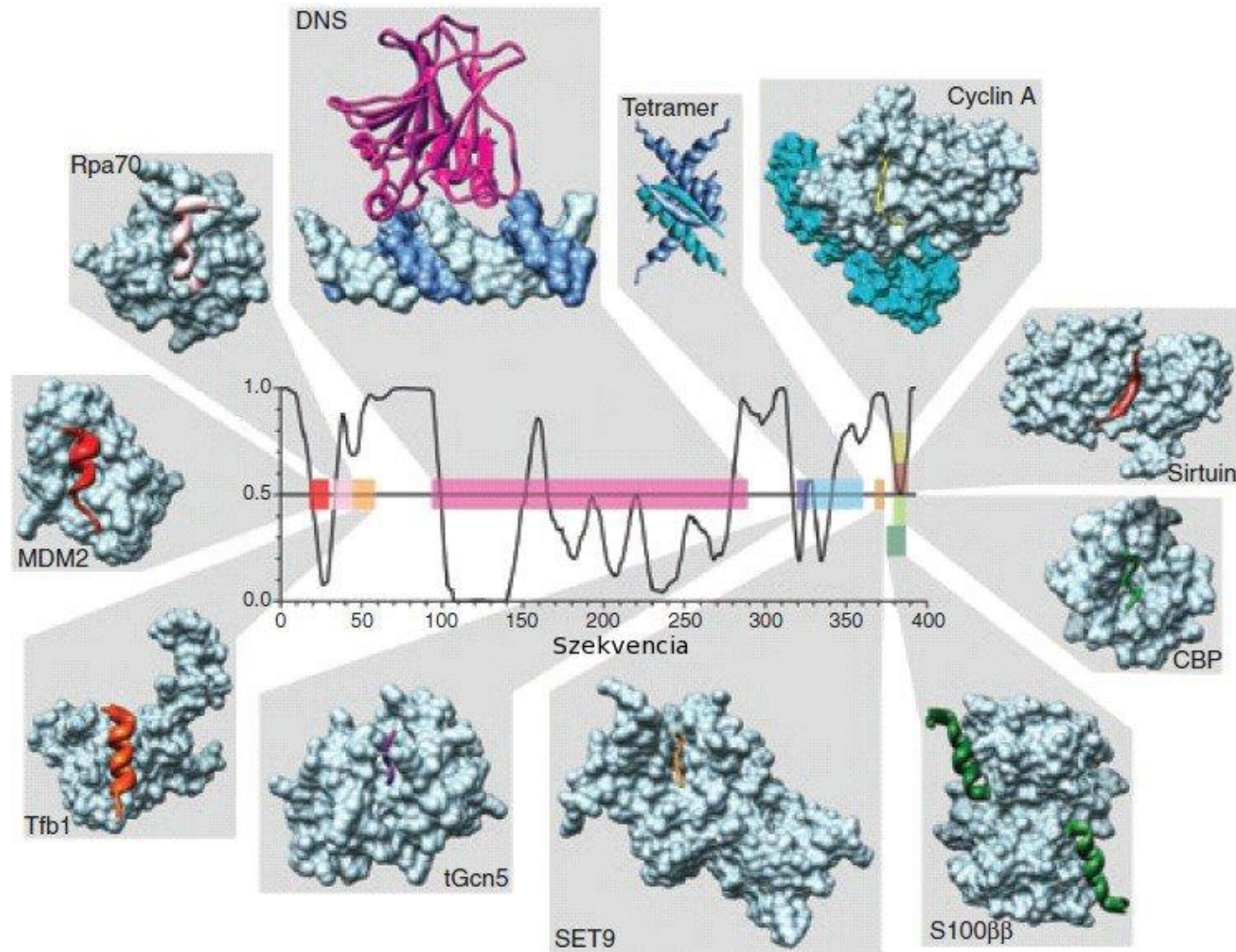
Intrinsically Disordered Proteins

- Exception to the Anfinsen dogma
- IDP ~ IUP
- lack of stable (steady) 3D structure
- highly dynamics structures
- usually many interaction partners
- different amino acid composition
usually with low complexity



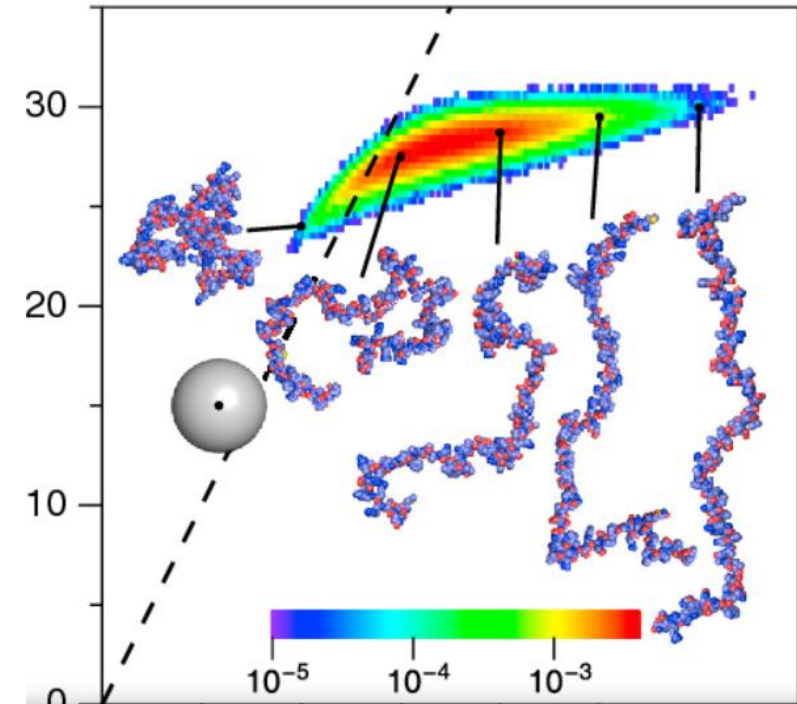
Cellular tumor antigene p53

regulator
of the cell
cycle and
apoptosis



Experimental validation

- Lack of secondary structure; CD spectroscopy
- Lack of X-ray electron density
- Radius of gyration



DisProt database



[Browse](#) [Release notes](#) [Download](#) [Help](#) [About](#)

[Login](#) [Feedback](#)



Version: **8.0**
Release: **2019_09**

Intrinsically disordered proteins

DisProt is a database of intrinsically disordered proteins. Disordered regions are manually curated from literature. DisProt annotations cover both structural and functional aspects of disorder detected by specific experimental methods. Annotation concepts and detection methods are encoded in the Disorder Ontology. Read [more about DisProt](#)

[Example 1](#) [Example 2](#)

Proteins per organism



H. sapiens: 578



M. musculus: 88



R. norvegicus: 50



S. cerevisiae: 128



E. coli: 7



A. thaliana: 33



D. melanogaster: 30



C. elegans: 13



Viruses: 126



Fungi: 156

Statistics

	Total	Not ambiguous
Proteins	1.6k	1.4k
Regions	3.5k	3k
Residues	164.1k	141.4k
Disorder content	19.7%	18.7%

Info

How to cite [Hatos A et al. DisProt: intrinsic protein disorder annotation in 2020](#)
Nucleic Acids Res., 2019. [\[NAR\]](#) [\[PubMed\]](#)

[Piovesan D et al. DisProt 7.0: a major update of the database of disordered proteins](#)
Nucleic Acids Res., 2016. [\[NAR\]](#) [\[PubMed\]](#)

API [REST API documentation](#) [here](#)

Integrated resources

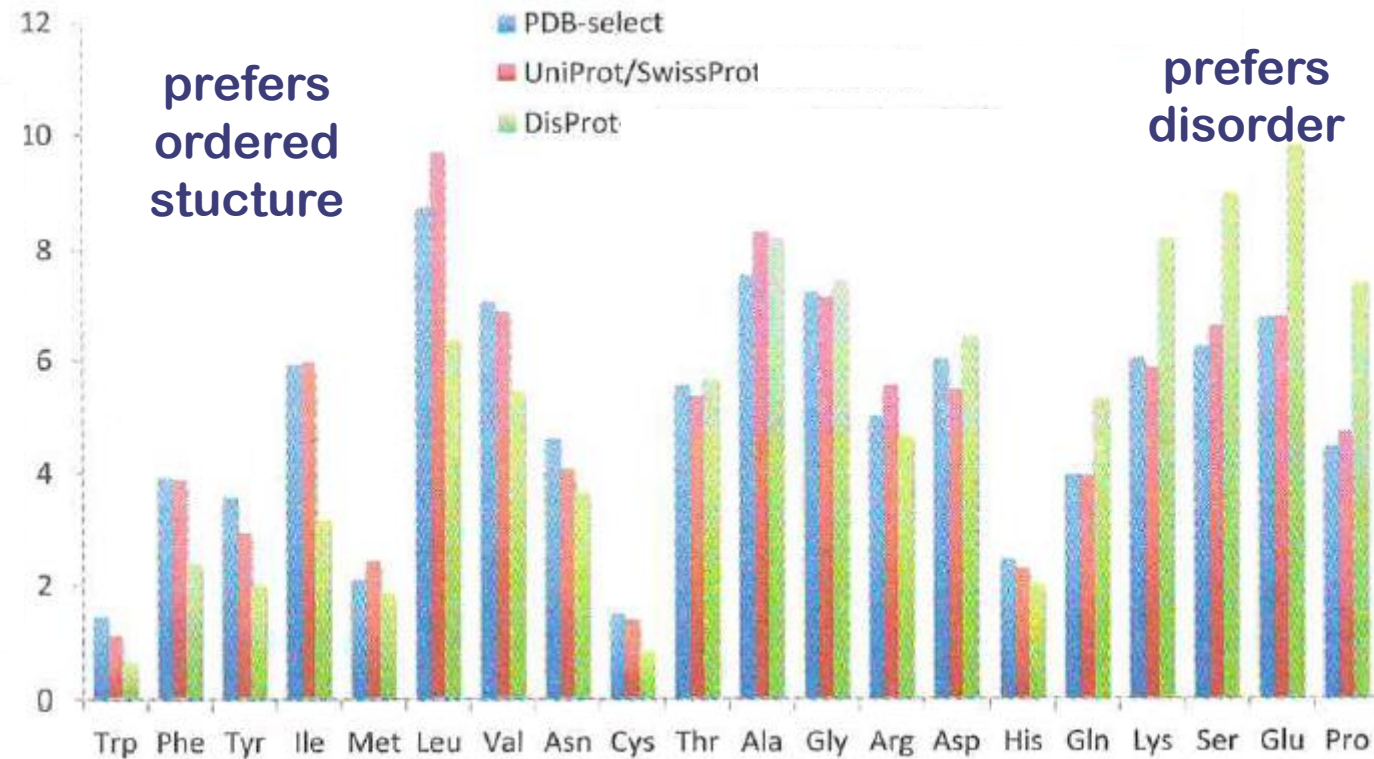


Order / disorder promoting residues

Scale	Order-promoting amino acid residues									
	W	F	Y	I	M	L	V	N	C	T
Top-IDP	-0.884	-0.697	-0.510	-0.486	-0.397	-0.326	-0.121	0.007	0.02	0.059
B-value	0.938	0.934	0.981	0.977	0.963	0.982	0.968	1.022	0.939	0.998
FoldUnfold	28.48	27.18	25.93	25.71	24.82	25.36	23.93	18.49	23.52	19.81
DisProt	-0.465	-0.381	-0.427	-0.393	0.197	-0.260	-0.302	-0.106	-0.546	-0.116

Scale	Disorder-promoting amino acid residues									
	A	G	R	D	H	Q	K	S	E	P
Top-IDP	0.06	0.166	0.180	0.192	0.303	0.318	0.586	0.341	0.736	0.987
B-value	0.994	1.018	1.026	1.022	0.967	1.041	1.029	1.025	1.052	1.050
FoldUnfold	19.89	17.11	21.03	17.41	21.72	19.23	18.19	17.67	17.46	17.43
DisProt	0.042	0.095	0.211	0.127	-0.127	0.381	0.370	0.201	0.469	0.419

Residue frequency



unfoldability by TOP-IDP scale → increasing

SLiMs: Short Linear Motifs

protein sequences mediating protein-protein interactions

linear, no real 3D structure

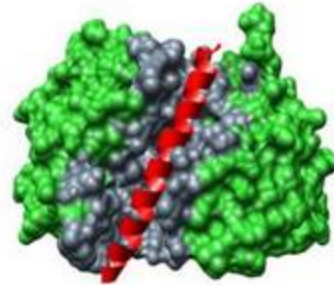
MoRFs

Molecular Recognition Features
motif-like peptides in disordered regions

disordered -> ordered

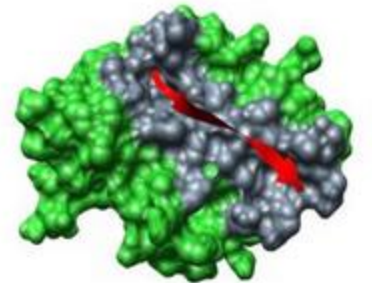
Molecular Recognition Features (MoRFs)

α -MoRF



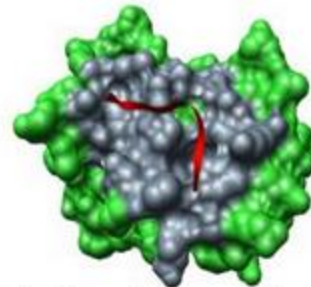
Proteinase A + Inhibitor
IA3

β -MoRF



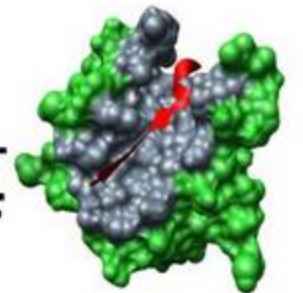
viral protein pVIc + Adenovirus 2
Proteinase

ι -MoRF



Amphiphysin + α -adaptin
C





complex-
MoRF

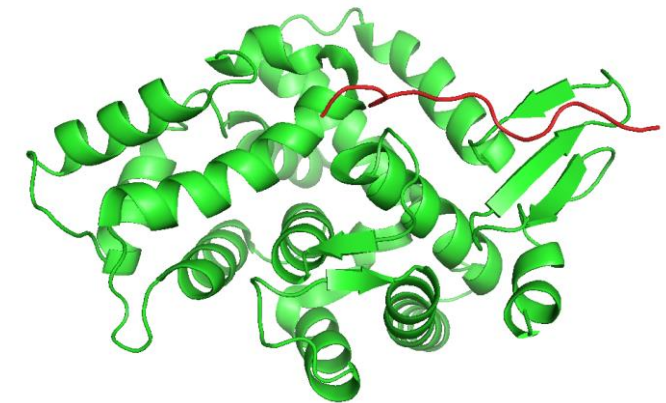
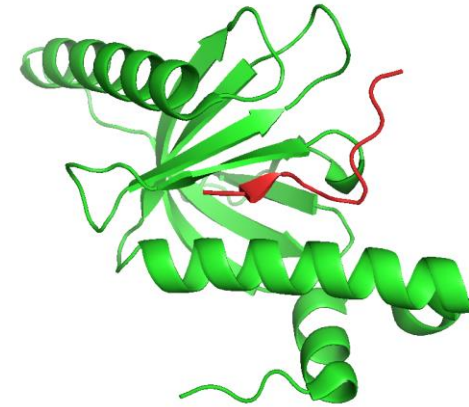


β -amyloid protein + protein
X11

ELMs

Eukaryotic Linear Motif resource

Accession	Acc. Gene-, Name	Start	End	Subsequence	Logic
 ELMI001563	 Q14114 LRP8 LRP8_HUMAN	860	867	KNTKSM <u>NFDNPVYR</u> KTTEEE	TP
Accession	Acc. Gene-, Name	Start	End	Subsequence	Logic
 ELMI001104	 Q14118 DAG1 DAG1_HUMAN	889	892	KGSRPKNMTPYRS <u>PPPY</u> VPP	TP



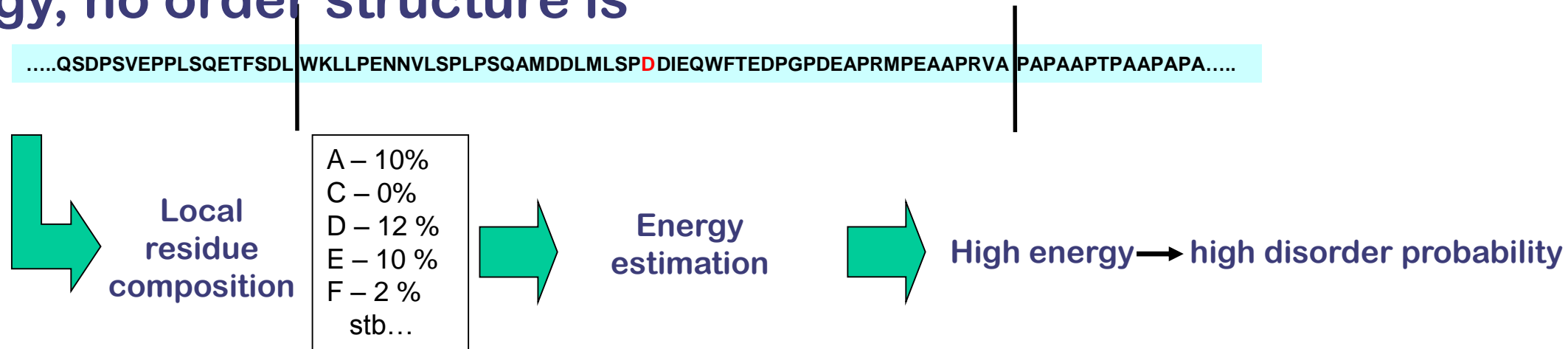
Prediction of disorder

- **Physics based approach**
 - IUPred
 - **Machine learning**
 - PONDR VSL2
 - DISOPRED3
 - Spritz
- Metaservers

The IUPred method

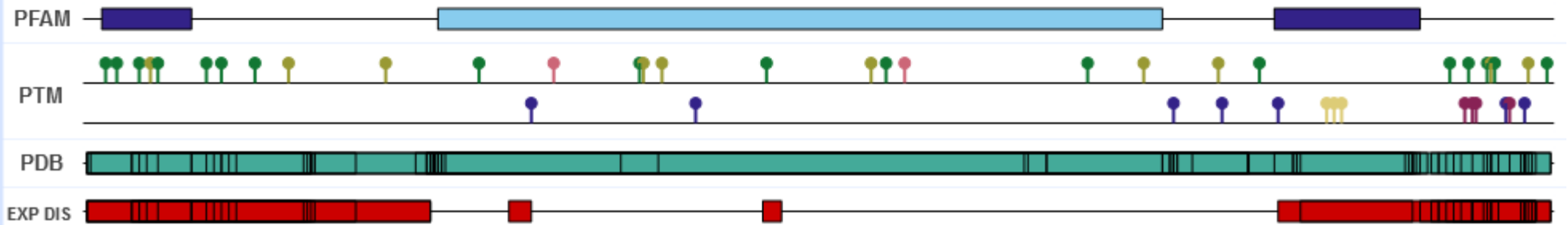
- Physics based approach using statistical pairwise potentials based on globular proteins
- transform observed frequencies of amino acid pair interactions into energy-like functions based on the Boltzmann statistics
- Idea: if the neighbour residues do not contribute to a low energy, no order structure is

A	– 7.67%
C	– 2.43%
D	– 4.92 %
E	– 5.43 %
F	– 3.19 %
...	



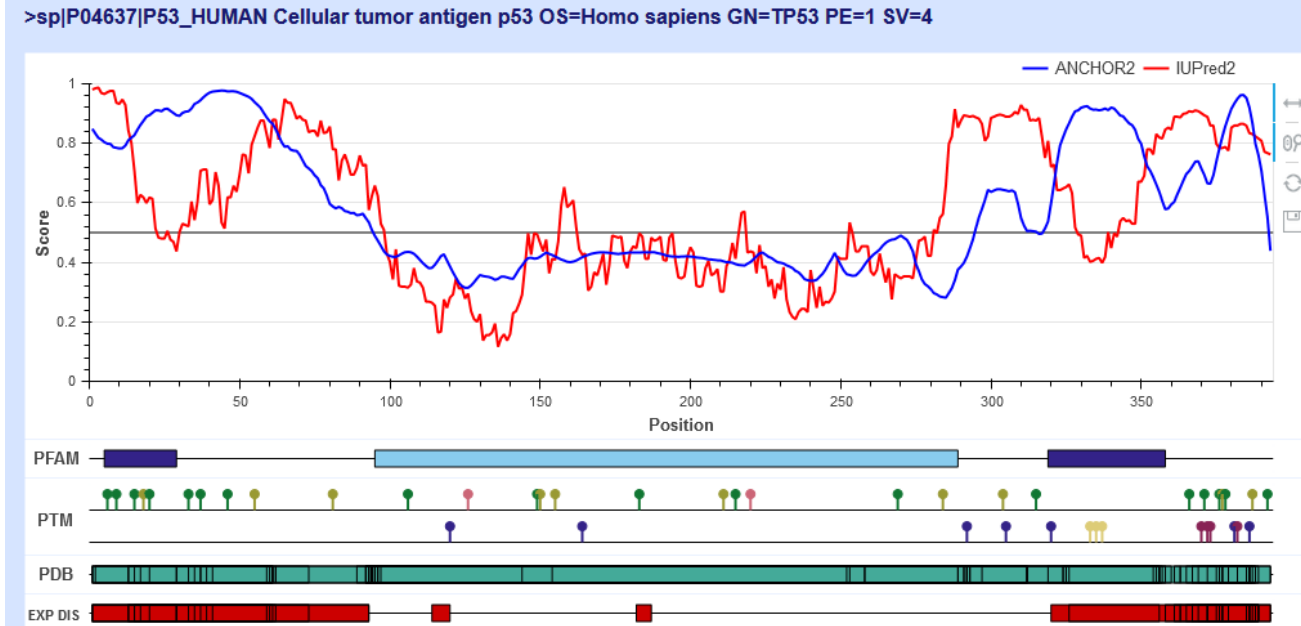
Output

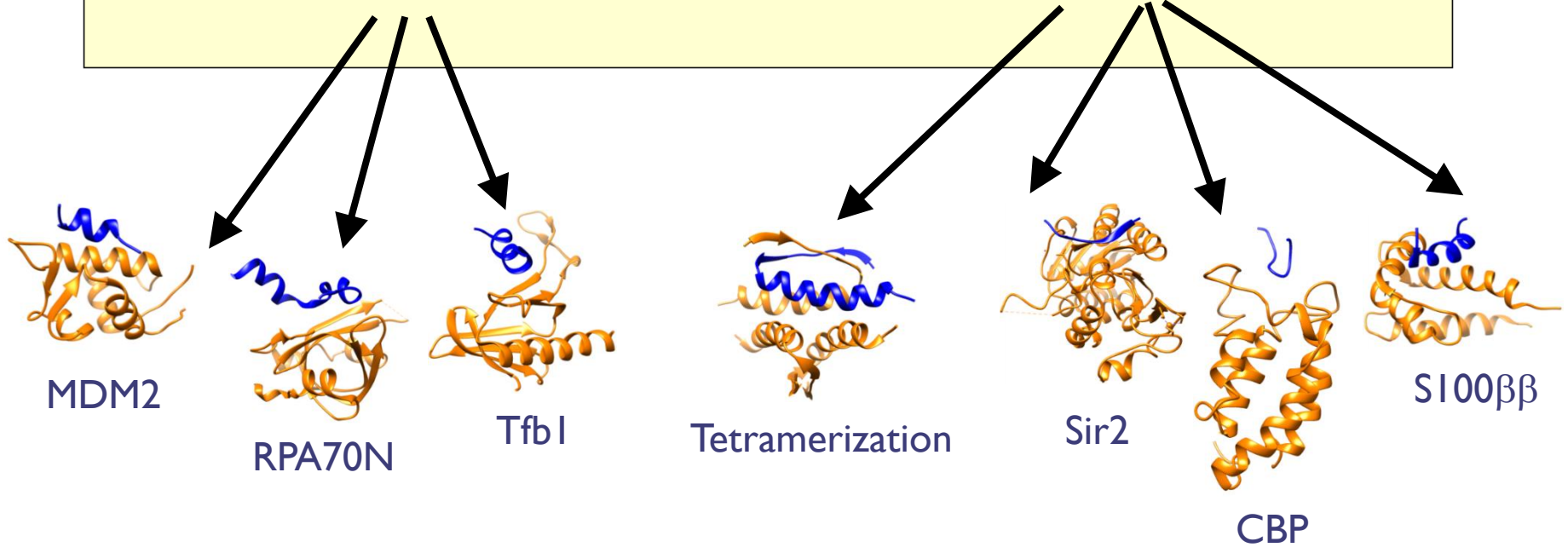
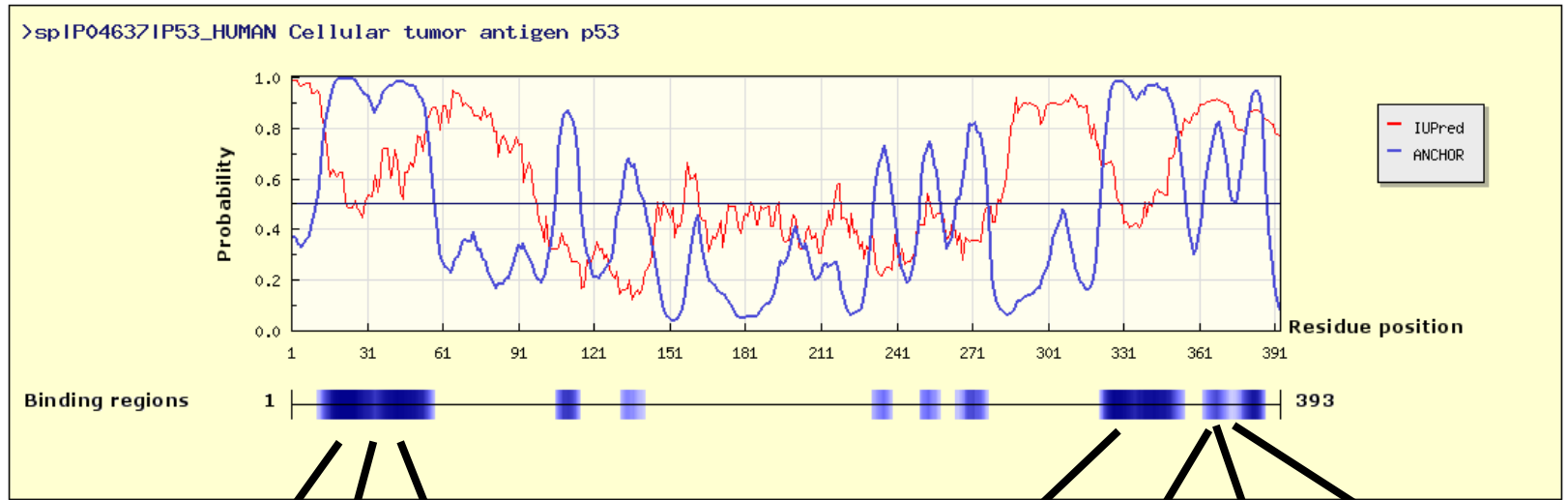
>sp|P04637|P53_HUMAN Cellular tumor antigen p53 OS=Homo sapiens GN=TP53 PE=1 SV=4



Anchor

- Prediction of disordered binding sites in globular proteins
- If the calculated energy is more favourable in a globular environment → binding site





Mutual Folding Induced by Binding

Mutual Folding Induced by Binding

MFIB

[Home](#) [Browser](#) [ProteinMap](#) [Search](#) [Downloads](#) [Statistics](#) [Help](#)

General Information

Function and Biology

Structure Summary

- Chain A
- Chain B

Evidence

Related Structure(s)

Structural Classification

Class:

- Other

Subclass:

- Ribbon-helix-helix (RHH)

General Information

Database accession: MF2140001

Name: Arc repressor

PDB ID: 1arq [PDB](#)

Experimental method: NMR

Assembly: homodimer

Source organism: *Enterobacteria phage P22*


Primer publication of the structure:

Bonvin AM, Vis H, Breg JN, Burgering MJ, Boelens R, Kaptein R
Nuclear magnetic resonance solution structure of the Arc repressor using relaxation matrix calculations. (1994) *J. Mol. Biol.* **236**: 328-41

PMID: 8107113 [PubMed](#)

Abstract:

The Arc repressor of Salmonella bacteriophage P22 is a dimeric sequence-specific DNA-binding protein. The solution structure of Arc has been determined from 2D NMR data using an "ensemble" iterative relaxation matrix approach (IRMA) followed by direct NOE refinement with DINOSAUR. A set of 51 structures was generated with distance geometry and further refined with a combination of restrained energy minimization and restrained molecular dynamics in a parallel refinement protocol. Distance constraints were obtained from an extensive set of NOE build-ups in H₂O and 2H₂O via relaxation matrix calculations from the ensemble of structures. Methyl group rotation, aromatic ring flaps and internal mobility effects (via order parameters obtained from a free molecular

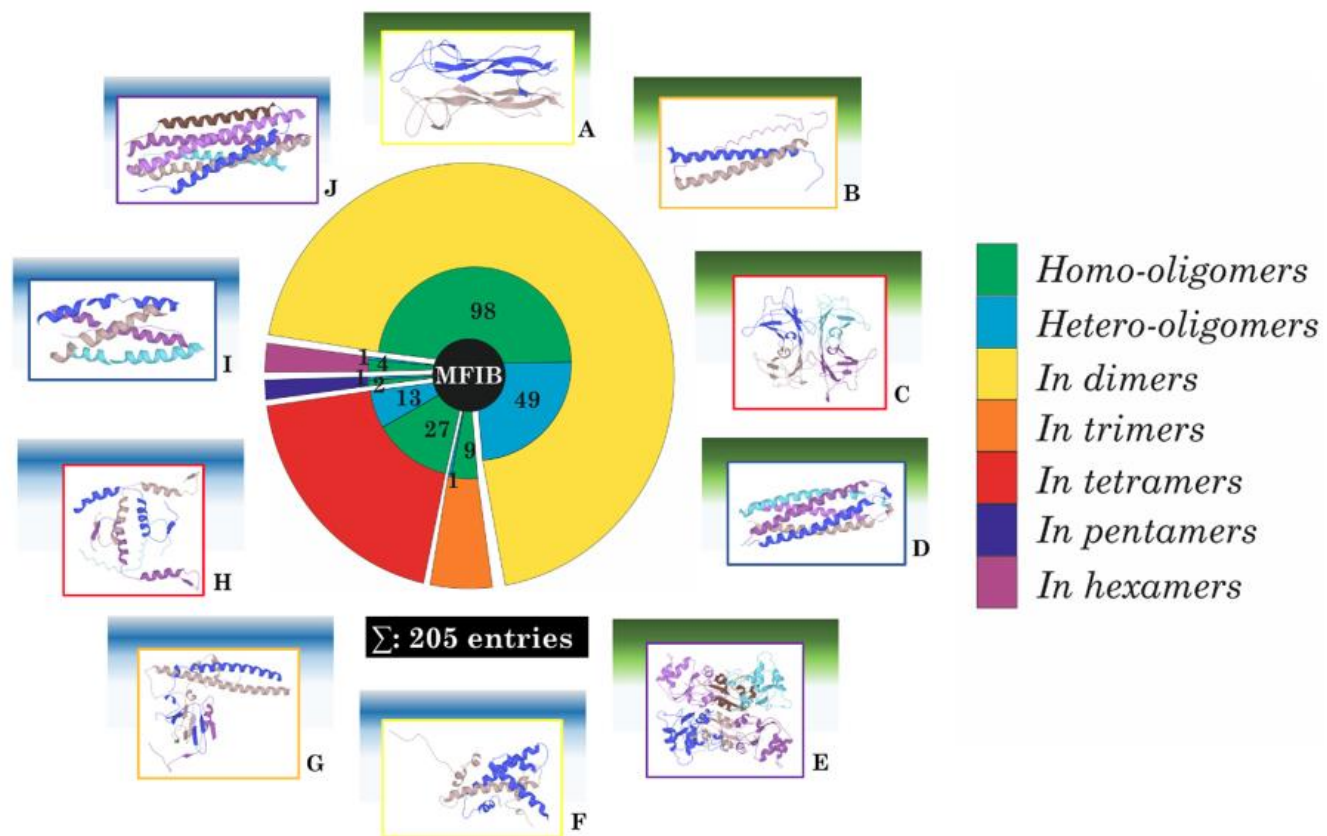
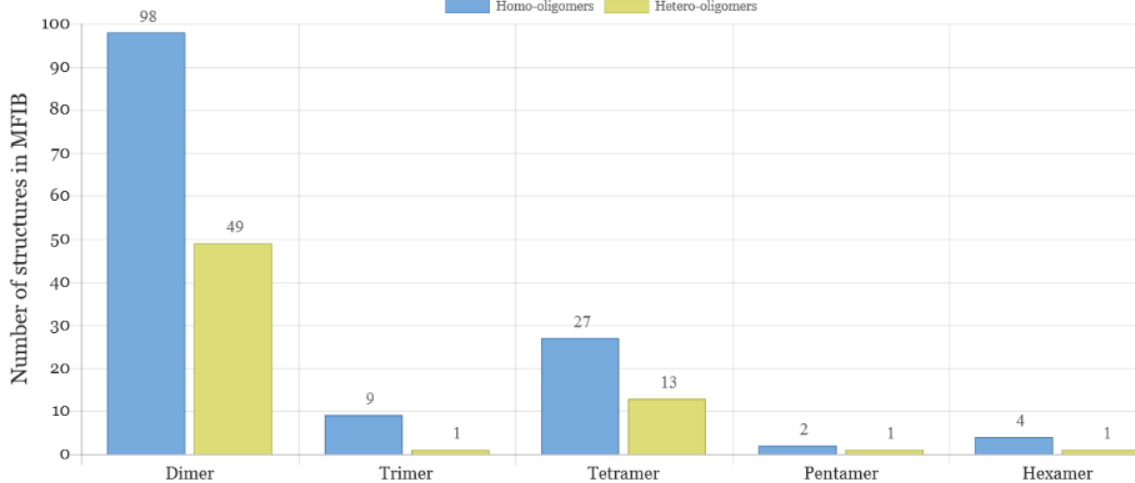


The molecule viewer shows the original PDB structure. Only the first NMR model was loaded.

Oligomeric states in MFIB

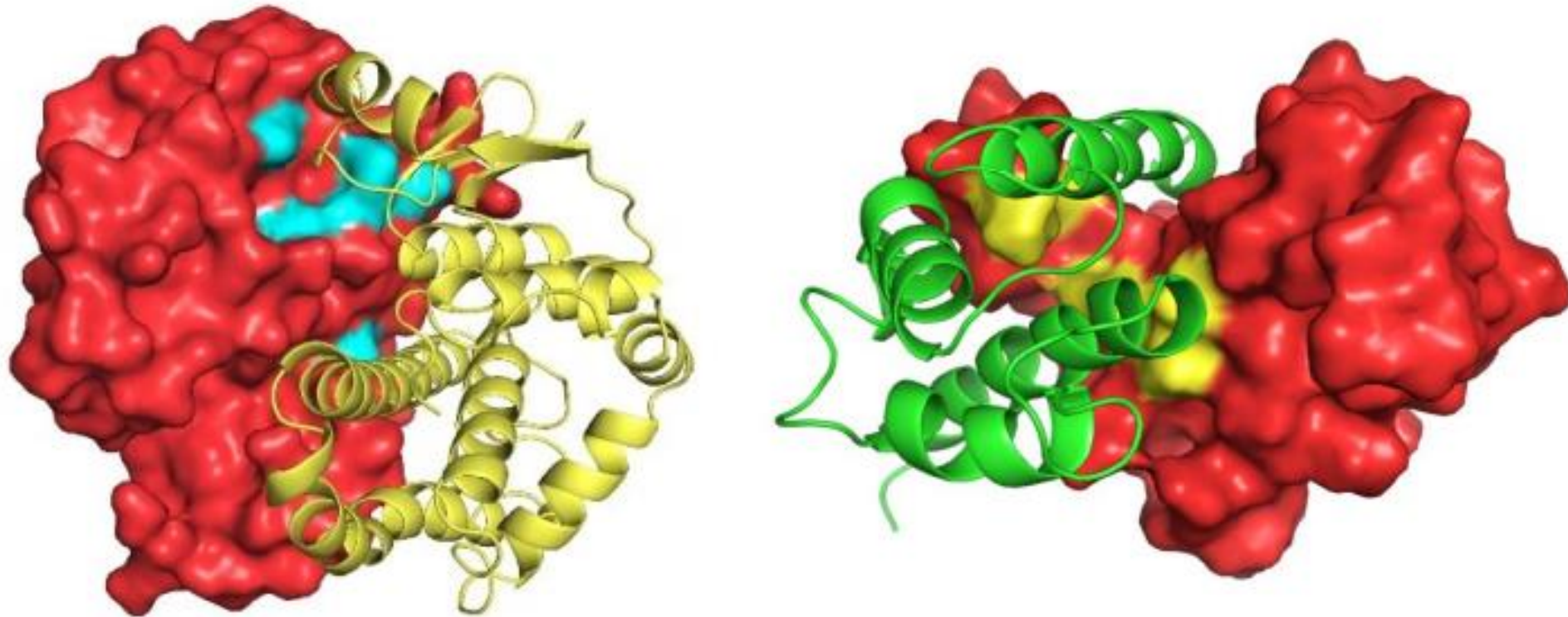
Oligomeric States in MFIB

■ Homo-oligomers ■ Hetero-oligomers

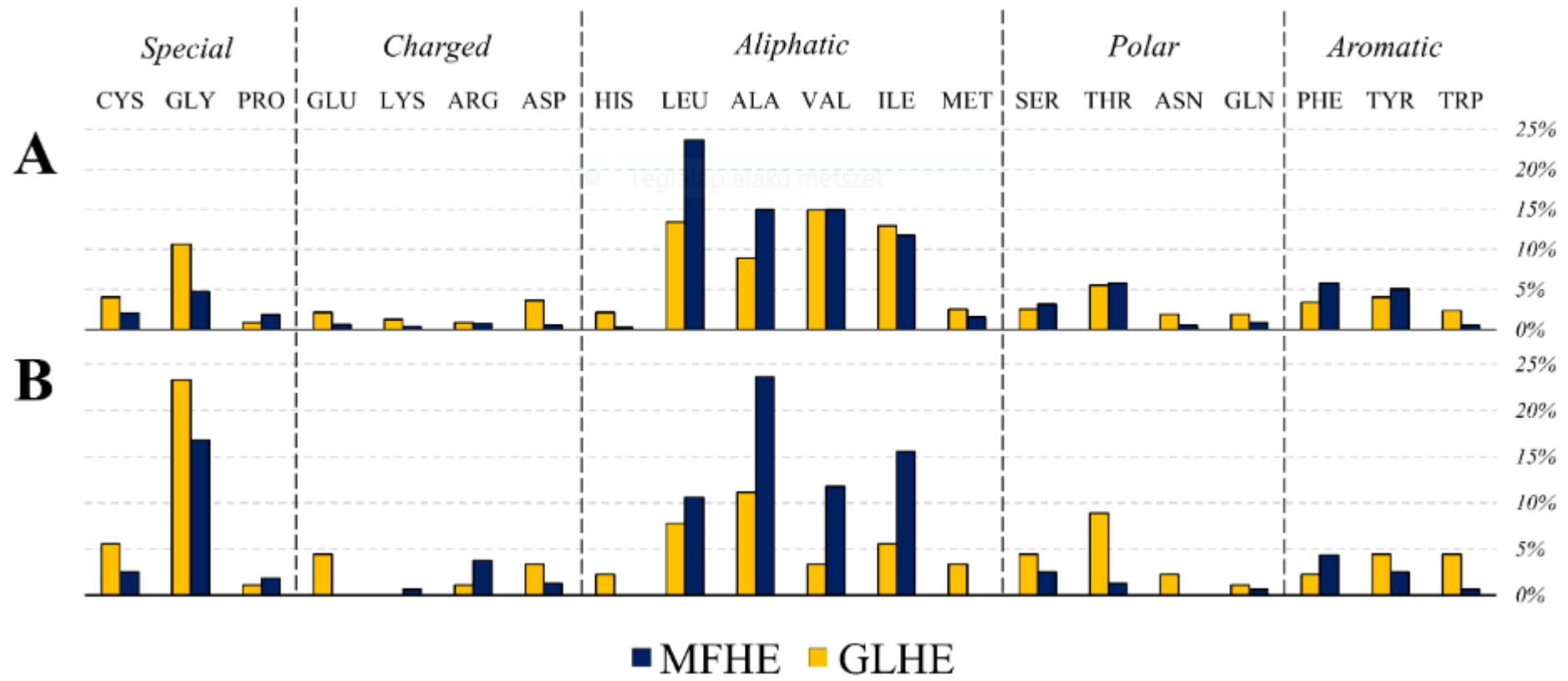


RSAMP

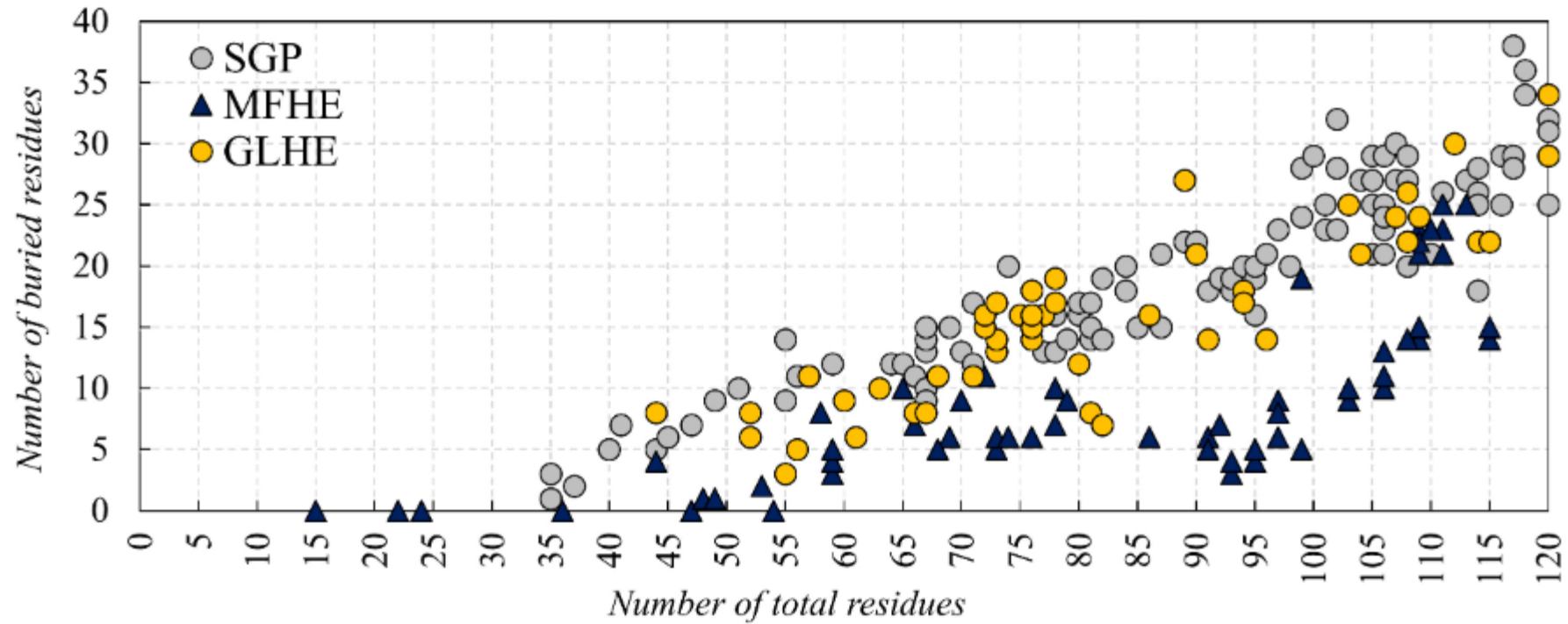
- Residue with Solvent Accessible Main-chain Patches



Amino acid compositions



Number of buried residues



Burial of residues in monomers / dimers

